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(21) International Application Number: PCT/US99/16417 (22) International Filing Date: 21 July 1999 (21.07.1999) (30) Priority Data: 09/181,601 29 October 1998 (29.10.1998) US 60/093,641 21 July 1998 (21.07.1998) US (60) Parent Application or Grant RUTGERS, THE STATE UNIVERSITY OF NEW JERSEY [/]; (). ANDERSON, Stephen [/]; (). MONTELLIONE, Gaetano [/]; (). HUANG, Yuanpeng [/]; (). ANDERSON, Stephen [/]; (). MONTELLIONE, Gaetano [/]; (). HUANG, Yuanpeng [/]; (). AUERBACH, Jeffrey, I. ; ().		Published
(54) Title: LINKING GENE SEQUENCE TO GENE FUNCTION BY THREE DIMENSIONAL (3D) PROTEIN STRUCTURE DETERMINATION (54) Titre: ETABLISSEMENT DE LIEN ENTRE UNE SEQUENCE DE GENE ET UNE FONCTION DE GENE PAR DETERMINATION DE LA STRUCTURE DE PROTEINE EN TROIS DIMENSIONS (3D) (57) Abstract <p>The present invention provides a structure-functional analysis engine for the high-throughput determination of the biochemical function of protein domains of unknown function, as exemplified in the flowchart of the figure. The present invention uses bioinformatics, molecular biology and nuclear magnetic resonance tools for the rapid and automated determination of the three-dimensional structures of proteins and protein domains.</p> (57) Abrégé <p>L'invention concerne un moteur d'analyse fonctionnelle de structure qui est destiné à la détermination à haut débit de la fonction biochimique de domaines de protéines dont la fonction est inconnue, selon l'exemple illustré dans l'organigramme de la figure. La présente invention utilise la bio-informatique, la biologie moléculaire et des instruments de résonance magnétique nucléaire à des fins de détermination rapide et automatisée des structures tridimensionnelles de protéines et de domaines de protéines.</p>		

Description

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TITLE OF THE INVENTION**LINKING GENE SEQUENCE TO GENE FUNCTION
BY THREE DIMENSIONAL (3D) PROTEIN
STRUCTURE DETERMINATION****5 CROSS-REFERENCE TO RELATED APPLICATIONS**

This application is a continuation-in-part of Provisional Patent Application No. 60/093,641 (filed July 21, 1998) and of U.S. Patent Application Serial No. 09/181,601 (filed October 29, 1998), which claims priority under 35 U.S.C. § 119(e) to Provisional Patent Application No. 60/063,679 (filed on October 29, 1997).

10 FIELD OF THE INVENTION

The present invention pertains to methods for elucidating the function of proteins and protein domains by examination of their three dimensional structure, and more specifically, to the use of bioinformatics, molecular biology, and nuclear magnetic resonance (NMR) tools to enable the rapid and automated determination of functions, as a means of genome analysis. The present invention further pertains to an integrated system for elucidating the function of proteins and protein domains by examining their three dimensional structure.

BACKGROUND OF THE INVENTION

One of the most powerful ways of identifying the biochemical and medical function of a gene product is to determine its three-dimensional structure. Although there are numerous examples in which the primary (i.e., linear) structure of a protein has provided key clues to its biochemical function, three dimensional (3D) structure determination is considered to be more definitive at establishing biochemical function. The process of elucidating the 3D structure of large molecules, such as proteins is generally thought of as slow and expensive.

In the past, most drugs were discovered by screening proprietary chemicals with animal models or receptor libraries. Today, this approach is being replaced by "combinatorial chemistry" and "rational drug design". These are the primary methods being used in the development of, for example, drugs targeted at the enzymes of the human AIDS virus.

5 What limits the drug discovery process today is not screening or medicinal
chemistry but the rate that the approximately 100,000 proteins in the human body can be
10 identified and prioritized as potential drug targets. Of particular significance for the
pharmaceutical industry are the emerging disciplines of bioinformatics and functional
5 genomics. Application of technologies developed in these areas will allow companies
to identify, in the next decade, the bulk of the most significant new drug targets. It has
been estimated that about 10,000 genes from the human genome are of potential value
15 in human medicine, but only a few percent of these genes have been isolated so far.
However, it is reported that by the year 2005 the raw sequence data for all of these genes
20 will have been determined by the Human Genome Project (HGP).

20 I. PROTEIN STRUCTURE

It is a generally accepted principle of biology that a protein's primary sequence
is the main determinant of its tertiary structure. Anfinsen, *Science* 181:223-230 (1973);
Anfinsen and Scheraga, *Adv. Prot. Chem.* 29:205-300 (1975); and Baldwin, *Ann. Rev.*
25 *Biochem.* 44:453-475 (1975). For over a decade, researchers have been studying the
theoretical and practical aspects of the folding of recombinant proteins.

For example, the "genetics" of protein folding using mutants of bovine
pancreatic trypsin inhibitor (BPTI) has been studied. Mutants of BPTI were prepared in
30 which several cysteine residues were replaced by alanine or threonine residues. These
20 mutants were then expressed in a heterologous *E. coli* expression system. Although
these mutants were found to fold into the proper conformation, the rate of the mutant
folding was somewhat slower than that exhibited by wild-type BPTI. Marks *et al.*,
Science 325:1370-1373 (1987).

35 Ma *et al.* have also studied the genetics of protein folding using mutants of
25 BPTI. Ma *et al.*, *Biochemistry* 36:3728-3736 (1997). The model system described by
Ma *et al.* predicts that a "rearrangement" mechanism to form buried disulfides at a late
stage in the folding reaction may be a common feature of redox folding pathways for
40 surface disulfide-containing proteins of high stability.

Nilsson *et al.* have reported that factors, such as peptidyl prolyl isomerase,
30 protein disulfide isomerase, thioredoxin, and Sec B, may interact with the unfolded
forms of specific classes of proteins, while members of the hsp70/DnaK and
hsp60/GroEL molecular chaperone families may play a more general role in protein
45 folding. Nilsson *et al.*, *Ann. Rev. Microbiol.* 45:607-635 (1991). Nilsson *et al.* further
disclose that intrinsic folding rates, or even translation rates, of nascent proteins may be

5 optimized by natural selection. Secretion, proteolysis and aggregation are other in vivo
processes that depend greatly in the folding behavior of a given protein. Thus, protein
folding involves an interplay between the intrinsic biophysical properties of a protein, in
10 both its folded and unfolded states, and various accessory proteins that aid in the
5 process.

Proteins are generally composed of one or more autonomously-folding units
known as domains. Kim *et al.*, *Ann. Rev Biochem.* 59:631-660 (1990); Nilsson *et al.*,
15 *Ann. Rev. Microbiol.* 45:607-635 (1991). Multidomain proteins in higher organisms are
encoded by genes containing multiple exons. Combinatorial shuffling of exons during
10 evolution has produced novel proteins with different domain arrangements having
different associated functions. This is thought to have greatly increased the ability of
higher organisms to respond to environmental challenges because, via recombinational
20 events, it has enabled genomes to readily add, subtract, or rearrange discrete
functionalities within a given protein. Patthy, *Cell* 41:657-663 (1985); Patthy, *Curr.*
15 *Opin. Struct. Bio.* 4:383-392 (1994); and Long *et al.*, *Science* 92:12495-12499 (1995).

25 II. INTERPRETATION OF A PROTEIN STRUCTURE

Several methods have been used to elucidate the 3D structure of a given protein
molecule. Chiefly, these methods are X-ray crystallography and Nuclear Magnetic
30 Resonance (NMR).

20 A. X-Ray Crystallography

X-ray crystallography is a technique that directly images molecules. A crystal of
the molecule to be visualized is exposed to a collimated beam of monochromatic X-rays
35 and the consequent diffraction pattern is recorded on a photographic film or by a
radiation counter. The intensities of the diffraction maxima are then used to construct
25 mathematically the three-dimensional image of the crystal structure. X-rays interact
almost exclusively with the electrons in the matter and not the nuclei.

The spacing of atoms in a crystal lattice can be determined by measuring the
angle and intensities at which a beam of X-rays of a given wave length is diffracted by
the electron shells surrounding the atoms. Operationally, there are several steps in X-
30 ray structural analysis. The amount of information obtained depends on the degree of
structural order in the sample. Blundell *et al.* provide an advanced treatment of the
45 principles of protein X-ray crystallography. Blundell *et al.*, *Protein Crystallography*,
Academic Press (1976), herein incorporated by reference. Likewise, Wyckoff *et al.*

provide a series of articles on the theory and practice of X-ray crystallography. Wyckoff *et al.* (Eds.), *Methods Enzymol.* 114: 330-386 (1985), herein incorporated by reference.

B. Nuclear Magnetic Resonance (NMR)

The classical approach for the analysis of NMR resonance assignments was first outlined by Wüthrich, Wagner and co-workers. Wüthrich, "NMR of proteins and nucleic acids" Wiley, New York, New York (1986); Wüthrich, *Science* 243:45-50 (1989); Billeter *et al.*, *J. Mol. Biol.* 155:321-346 (1982), all of which are herein incorporated by reference. For a general review of protein determination in solution by nuclear magnetic resonance spectroscopy, see Wüthrich, *Science* 243:45-50 (1989). See also, Billeter *et al.*, *J. Mol. Biol.* 155:321-346 (1982).

Wüthrich's classical approach can be briefly summarized in the following seven steps:

- Step 1: Identification of individual resonances associated with each spin system, and designation of key atom types (e.g., H^N , H^α , N , C^α , C^β , etc.).
- Step 2: Classification of each identified spin system with respect to one or more possible amino acid residue type(s).
- Step 3: Identification of possible sequential relations between spin systems using inter-residue NOESY or triple-resonance data.
- Step 4: Unique mapping of strings of sequentially-connected spin systems to segments of the amino acid sequence, thus establishing "sequence specific assignments."
- Step 5: Extension of assignments to resonances of peripheral side-chain nuclei in each spin system, and determination of stereospecific assignments.
- Step 6: Generation of distance constraints using assigned resonance frequencies to interpret NOESY, scalar-coupling, and hydrogen/deuterium-exchange data in terms of "sequence-specific distance constraints."
- Step 7: Structure generation using these constraints.

Automated implementation of these methods have made use of exhaustive search, constraint satisfaction, heuristic best-fit or branch-and-bound limited search, genetic, neural net, pseudoenergy minimization, and simulated annealing satisfaction. Billeter *et al.*, *J. Magn. Resonance* 76:400-415 (1988); Zimmerman *et al.*, In:

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Proceedings of the First International Conference of Intelligent Systems for Molecular Biology. Washington: AAAS Press (1994); Zimmerman *et al.*, *J. Biomol. NMR* 4:241-256 (1994); Zimmerman *et al.*, *Curr. Opin. Struct. Bio.* 5:664-673 (1995); and Zimmerman *et al.*, *J. Mol. Bio.* 269:592-610 (1997).

5 Under traditional methodology, before a given protein is studied at the 3D level, the researcher had already obtained detailed experimental information regarding the protein's function and characteristics. The 3D structure is typically the last of many experiments performed over many years of study. The 3D structure information is then used to refine the researcher's understanding of the given protein. Thus, under traditional methodology, it is very rare that the 3D structure of a given protein is determined before its biochemical function has been determined by other methods.

10 The present invention represents a paradigm shift in methodology because the researcher would first determine the 3D structure of a protein of unknown function and then use this structure to gain clues as to its function, which would be subsequently validated by appropriate biochemical assays.

SUMMARY OF THE INVENTION

5 The present invention describes an integrated system for rapid determination of the three-dimensional structures of proteins and protein domains and application of this technology in a high-throughput analysis of human and other genomes for drug discovery purposes.

10 The "structure-function analysis engine" described herein has the potential to discover the functions of novel genes identified in the human and other genomes faster than existing genetic or purely computational bioinformatics methods.

15 The present invention employs:

- 20 1. Bioinformatics methods, including the analysis of exon-exon phases and other methods for segmenting or "parsing" DNA sequences of novel genes into domain-encoding regions;
2. Robust and general "domain trapping" methods for producing correctly-folded recombinant protein domains of novel biomedically-important human disease gene products;
3. Robust and general methods for high level expression and isotopic enrichment of these domains for NMR and X-ray crystallographic studies;

4. Screening methods to identify protein domain constructs that exhibit the properties required for structural analysis by NMR or X-ray crystallography;
5. Computer software, NMR pulse sequences, and related NMR technologies that provide fully automated analysis of protein structures from NMR data;
6. NMR spectroscopy methods for determining 3D structures of these domains;
7. Improved methods for mapping new domain structures to proteins in the Protein Data Bank that have similar structures and biochemical functions;
8. A relational data base of the empirical properties of expressed domains for organizing and integrating the biophysical and biological information derived from these studies, as well as methods for making such relational data bases; and
9. A method for integrating all of the above into a large-scale, high-throughput macromolecular "structure-function analysis engine," and the application this "structure-function analysis engine" to the discovery of biochemical functions of hundreds of genes from humans and human pathogens.

The specific biomedical gene targets that this technology can be used to develop include:

1. Domains from the human Alzheimer's β peptide precursor protein (APP).
2. Domains from other proteins genetically implicated in neoplastic, metabolic, neurodegenerative, cardiovascular, psychiatric and inflammatory disorders.
3. Domains from proteins associated with infectious agents (e.g., bacteria, fungi and viruses).

The present invention provides a high-throughput method for determining a biochemical function of a protein or polypeptide domain of unknown function comprising: (A) identifying a putative polypeptide domain that properly folds into a stable polypeptide domain, the stable polypeptide having a defined three dimensional structure; (B) determining three dimensional structure of the stable polypeptide domain; (C) comparing the determined three dimensional structure of the stable polypeptide

5 domain to known three-dimensional structures in a protein data bank, wherein the
comparison identifies known structures within the protein data bank that are
homologous to the determined three dimensional structure; and (D) correlating a
10 biochemical function corresponding to the identified homologous structure to a
5 biochemical function for the stable polypeptide domain.

The present invention further provides an integrated system for rapid
determination of a biochemical function of a protein or protein domain of unknown
15 function: (A) a first computer algorithm capable of parsing the target polynucleotide
into at least one putative domain encoding region; (B) a designated lab for expressing
10 the putative domain; (C) an NMR spectrometer for determining individual spin
resonances of amino acids of the putative domain; (D) a data collection device capable
of collecting NMR spectral data, wherein the data collection device is operatively
20 coupled to the NMR spectrometer; (E) at least one computer; (F) a second computer
algorithm capable of assigning individual spin resonances to individual amino acids of a
15 polypeptide; (G) a third computer algorithm capable of determining tertiary structure of
a polypeptide, wherein the polypeptide has had resonances assigned to individual amino
acids of the polypeptide; (H) a database, wherein stored within the database is
25 information about the structure and function of known proteins and determined proteins;
and (I) a fourth computer algorithm capable of determining 3D structure homology
20 between the determined three-dimensional structure of a polypeptide of unknown
function to three-dimensional structure of a protein of known function, wherein the
protein of known structure is stored within the protein database.

The present invention further provides a high-throughput method for
determining a biochemical function of a polypeptide of unknown function encoded by a
35 target polynucleotide comprising the steps: (A) identifying at least one putative
polypeptide domain encoding region of the target polynucleotide ("parsing"); (B)
expressing the putative polypeptide domain; (C) determining whether the expressed
putative polypeptide domain forms a stable polypeptide domain having a defined three
40 dimensional structure ("trapping"); (D) determining the three dimensional structure of
30 the stable polypeptide domain; (E) comparing the determined three dimensional
structure of the stable polypeptide domain to known three dimensional structures in a
Protein Data Bank to determine whether any such known structures are homologous to
the determined structure; and (F) correlating a biochemical function corresponding to
45 the homologous structure to a biochemical function for the stable polypeptide domain.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 provides a flow chart of the high-throughput structure/function analysis system of the present invention.

Figure 2A provides the far UV circular dichroism spectra of the purified recombinant APP NTD2-3 domain. Figure 2B provides the near UV circular dichroism spectra of the purified recombinant APP NTD2-3 domain.

Figure 3 provides a NMR spectra of the purified recombinant APP NTD2-3.

Figure 4 provides a hydrogen-deuterium exchange time course for the purified recombinant APP NTD2-3.

Figure 5 provides the results of a cooperative thermal unfolding experiment of the purified recombinant APP NTD2-3.

Figure 6 provides the results of the NMR ^{15}N - ^1H heteronuclear single quantum coherence (HSQC) spectral analysis of the NTD2-3 domain collected on a Varian Unity 500 spectrometer.

Figure 7 provides the 2D ^{15}N - ^1H HSQC spectrum of CspA at pH 6.0 and 30°C.

Figure 8A provides an illustration of information derived from triple resonance data sets used for establishing intraresidue and sequential correlations of spin systems.

Figure 8B provides an illustration of NMR data used to identify structural elements in CspA. Slowly exchanging backbone amides ($t_{1/2} > 3$ min at pH 6.0 and 30°C) are indicated by filled circles ($t_{1/2} < 30$ min) or stars ($t_{1/2} > 30$ min.). Values of $^3J(\text{H}^{\text{N}}-\text{H}^{\alpha})$ coupling constants are indicated by vertical bars; filled bars indicate that the data provided a useful estimate ($\pm 0.5\text{Hz}$) of the corresponding coupling constant, while open bars indicate that the experimental data provide only an upper bound on its value. Values of conformation-dependent secondary shifts $\Delta\delta\text{C}^{\alpha}$ and $\Delta\delta\text{C}^{\beta}$ are plotted with solid bars. The locations of the five β -strands are indicated with arrows.

Figure 9 provides a flow chart of a NOESY_ASSIGN Process of the present invention.

Figures 10A and B provide the 3D structure of the Zdom protein.

Figures 11, 12 and 13 provide results of an automated assignment analysis for the Zdom protein.

Figures 14, 15 and 16 provide results of a manual assignment analysis for the Zdom protein.

Figure 17 provides the 3D structure of the Cspa protein.

Figures 18, 19 and 20 provide results of an automated assignment analysis for the Cspa protein.

Figures 21, 22, and 23 provide results of a manual assignment analysis for the Cspa protein.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

One of the best clues to a protein's function is its structure. The present invention describes a structure-based bioinformatics platform to be used in "functional genomics" analyses of the torrent of DNA sequence data emerging from the international HGP. This technology will allow for the isolation of novel biopharmaceuticals and/or drug targets from gene sequence information with an efficiency that is far beyond present day capabilities. By developing extremely fast yet rigorous technologies for macromolecular structure determination, it is possible to convert the stream of one-dimensional DNA sequence information emerging from human genome research efforts into 3D protein structures. This 3D structural information can then be used to map these human gene products to protein families with similar biochemical functions.

The present invention describes a "drug discovery search engine" that allows human genetic and genomic data to be smoothly interfaced with proven rational drug design and combinatorial chemistry approaches. The technology described herein enables determination of the structures for virtually the entire complement of human protein domains, encoded in the approximately 100,000 human genes.

I. STRUCTURE SUGGESTS FUNCTION

It is a tenet of modern structural biology that structure suggests function: a given protein "fold" tends to be used over and over again in nature for a restricted set of biological functions. Knowledge of the structure of a new protein often reveals kinship to a family of other proteins with already known functions, and thus provides strong clues regarding the biochemical function of the protein at hand. Holm *et al.*, *Science* 273:595-603 (1996); Bork *et al.*, *Curr. Opin. Struct. Bio.* 4:393-403 (1994); Brenner *et al.*, *Proc. Natl. Acad. Sci. (U.S.A.)* 95:6073-6078 (1998), all of which are herein incorporated by reference. This kinship relationship is a natural manifestation of the fact that families of protein molecules have evolved from a common ancestral molecule, and that in the course of this evolution the 3D structure is largely preserved while new, though chemically related, biochemical functions are adopted. This is precisely the reasoning behind the assigning of "expressed sequence tag" (EST) sequences to known protein families using one-dimensional sequence comparisons.

Evolution generally acts to conserve 3D structures rather than the amino acid sequences of proteins. For this reason, proteins have often evolved over time so that their sequences exhibit no obvious similarity while their structures remain highly homologous. In practical terms, this means that simple sequence comparisons overlook many -- and perhaps even most -- instances of protein-protein relatedness. However, this relatedness, with all of its functional implications, can easily be identified by 3D structure comparisons.

The multidomain nature of many mammalian proteins makes them more difficult to express in recombinant form and also impedes their structure determination by X-ray crystallography or NMR. The expression and structure determination of an isolated domain is, in contrast, less problematical. Since an isolated domain comprises one or more discrete functional units in a protein, knowing structure-function information about a given individual domain in a multicomponent protein generally provides key information that can be used to proceed with drug development on the full-length protein. The "domain trapping" methods of the present invention generate many novel gene products suitable for structural analysis by NMR spectroscopy and X-ray crystallography.

Recent developments in the areas of high-level protein expression technology, X-ray crystallography, heteronuclear NMR spectroscopy, and artificial intelligence (AI)-based structural analysis software, have dramatically improved the speed and lowered the cost of protein structure determination. Estimates of the total number of human genes in the genome (approximately 10^5) contrast dramatically with estimates of the total number of protein folds in nature (approximately 10^3), and it has been estimated that one-third to one-half of these folds have already been described. Chothia *et al.*, *Nature* 357:543-544 (1992). Simple statistics imply that many new gene products will exhibit structures that map to existing fold classes associated with proteins of known biochemical function. Thus, the harvest of functional information about new human genes from this approach will be immediate.

II. DESIGN OF A HIGH-THROUGHPUT SYSTEM FOR DETERMINING PROTEIN STRUCTURES AND FUNCTIONS

Figure 1 provides a flow chart of the high-throughput structure/function analysis used in the present invention for analyzing human and pathogen gene products. This flow chart outlines the general methods of the present invention. Each sub-step of the

present invention is outlined in detail below. It is to be understood that the hardware disclosed herein can be or is operatively linked to one or more computers.

A. Approaches For Identifying Novel Protein Domains

The present invention provides a method for predicting the location of domains and domain boundaries within a given DNA sequence. Under one embodiment, this is accomplished through a knowledge based application which segments or "parses" genomic or cDNA sequences of genes into domain encoding sequences. Under another embodiment, the knowledge based application of the present invention can also segment or "parse" mRNA sequences into domain encoding sequences. Preferably, the knowledge based application of the present invention is encoded within a computer algorithm software application. Preferably, this expert system applies rules developed on a set of experimentally-verified DNA sequence/protein domain comparisons that have been compiled from public sequence and protein structure databases. Thus, for a novel gene sequence, this expert system generates the predicted domains and/or domain boundaries which are then used to create domain-specific expression constructs.

Under one of the preferred embodiments, the gene sequence is parsed by the exon phase rule. Exon termini (5'- or 3') that begin or end within protein coding regions can be classified according to their "phase": an exon terminus that falls between two codons is called a "phase 0" terminus; an exon terminus that starts or stops after the first nucleotide in the codon is called a "phase 1" terminus; and an exon terminus that starts or stops after the second nucleotide in the codon is called a "phase 2" terminus. For example, where ("*") marks the positions of an exon-exon junction--

Phase 0: *
5' ... -A-T-G-G-G-A-C-T-C- ... 3'
... - Met - Gly - Leu - ...

Phase 1: *
5' ... -A-T-G-G-G-A-C-T-C- ... 3'
... - Met - Gly - Leu - ...

Phase 2: *
5' ... -A-T-G-G-G-A-C-T-C- ... 3'
... - Met - Gly - Leu - ...

5 The genetic coding sequences for protein domains, which have been reported to
have been "shuffled" between various genes during evolution, should be bounded by
exon termini of the same phase (or by the N- or C-terminal ends of the holoprotein),
10 otherwise insertion of these domains into a host gene would result in a frame-shift
5 mutation in the downstream sequences upon splicing (Patthy, *Cell* 41:657-663 (1985);
Patthy, *FEBS Letters* 214:1-7 (1987); Patthy, *Cur. Opin. Struct. Bio.* 4:383-392 (1994),
all of which are herein incorporated by reference). Therefore, the domain encoding
15 regions should be bounded on both sides by phase 0 exon termini, by phase 1 exon
termini, or by phase 2 exon termini, but not by termini of different phases.

10 As part of the mechanism of molecular evolution, structural and functional
domains are mixed and matched between protein sequences through the processes of
gene duplication and crossover. Accordingly, under one preferred embodiment domains
20 are identified by looking for segments of gene sequences that are conserved across many
genes from different organisms. Known domain families generally involve 50 - 300
15 amino-acid long segments that are observed as portions of many different proteins.
Bioinformatics algorithms capable of identifying these conserved segments, or gene-
25 fragment clusters, in the data base of gene sequences have been reported. These
algorithms can be used to identify candidate domain-encoding regions in novel gene
sequences. Gouzey *et al.*, *Trends Biochem. Sci.* 21:493 (1994), herein incorporated by
20 reference.

30 Under a second preferred embodiment, domains from gene sequence data are
identified through predictions of their interdomain boundaries. There is ample evidence
from molecular evolution and cell biology studies that information regarding domain
boundaries is embedded in the sequences of protein coding genes. Some reports have
25 claimed that rare codon clusters, which cause ribosomal pausing during translation, are
35 correlated with domain boundaries. Purvis *et al.*, *J. Mol. Biol.* 193:413-417 (1987);
Nilsson *et al.*, *Ann. Rev. Microbiol.* 45:607-635 (1991); Thanaraj *et al.*, *Protein Sci.*
5:1973-1983 (1996); Thanaraj *et al.*, *Protein Sci.* 5:1594-1612 (1996); and Guisez *et al.*,
40 *J. Theor. Biol.* 162:243-252 (1993), all of which are herein incorporated by reference.
30 Messenger RNA secondary structure have also been reported to play such a
"punctuation" role during translation.

One embodiment of the present invention employs an algorithm that identifies
such sequence features and compares these data with the actual domain sequences in the
45 relational database of the present invention. The relational database of the present
35 invention contains domain sequence information of known and determined protein
domains. It is understood that the relational database of the present invention will

expand over time such that each polypeptide domain determined using the methods of the present invention will be added to the relational database. Under this embodiment, it is possible to rigorously assess the reliability of these bioinformatics methods of domain prediction and, iteratively, modify the software to improve its reliability. Neural nets and genetic algorithms both can be used for deriving rules for domain boundaries from this knowledge base. This invention markedly accelerates productivity by greatly reducing the number of expression constructs that would have to be tested in order to correctly parse a novel gene sequence into its component domain sequences.

Under another embodiment, the solution structure of a protein or protein domain can be analyzed by a method that combines enzymatic proteolysis and matrix assisted laser desorption ionization mass spectrometry (Cohen *et al.*, *Protein Sci.* 4:1088-1099 (1995), Seielstad *et al.*, *Biochem.* 34:12605-12615 (1995), both of which are incorporated by reference in their entirety). This method is capable of inferring structural information from determinations of protection against enzymatic proteolysis as governed by solvent accessibility and protein flexibility. Preferably, the proteolytic enzymes employed by this method include trypsin, chymotrypsin, thermolysin, and ASP-N endoprotease.

B. "Domain Trapping": Expression And Biophysical Characterization Of Putative Recombinant Protein Domains

With respect to genes of unknown function, the investigator, generally, does not have available an enzyme assay or other obvious activity-based means to assess the biochemical activity of a novel recombinant protein domain. The present invention has addresses this difficulty in a three-pronged manner. First, the present invention uses a reliable and high yield expression system for protein expression. For example, a secretion-based protein A fusion system that is one of the most tested and reliable methods known for producing correctly-folded recombinant proteins in the *E. coli* periplasm. Nilsson *et al.*, *Methods Enzymol.* 185:144-161 (1990), herein incorporated by reference. Alternatively, the pET plasmid expression system may be used. Studier *et al.*, *J. Mol. Bio.* 189:113-130 (1986), herein incorporated by reference. Second, the present invention uses a set of activity-independent biophysical criteria to assess whether the protein domain has properly folded. This set of criteria has been developed through extensive study of recombinantly-expressed protein folding mutants. Finally, based on the supposition that autonomous folding of the protein domain can be prevented due to too much or too little polypeptide sequence information, respectively,

(Kim *et al.*, *Ann. Rev. Biochem.* 59:631-660 (1990); Nilsson *et al.*, *Ann. Rev. Microbiol.* 45:607-635 (1991), both of which are herein incorporated by reference), the present invention uses systematic strategies for identifying and trapping domains that enables it to use a combination of molecular biological and biophysical methods to experimentally parse any gene into its component domains. In other words, a polypeptide domain has a "defined three dimensional structure" when that polypeptide domain exhibits the activity-independent biophysical criteria of a properly folded domain.

Under one preferred embodiment, an activity-independent biophysical criteria used to assess the correctness of folding of a protein includes circular dichroism measurements. More preferably, characterization of an isolated domain of a protein is analyzed by circular dichroism measurements in the far UV. An ellipticity minimum at 222 nm is indicative of α -helical secondary structure. Preferably, CD measurements at longer wavelengths are also determined (for a general review of CD and other methods, see Creighton, *Proteins: Structure and molecular properties*, 2nd Ed., W. H. Freeman & Co., New York, New York (1993, and related texts), herein incorporated by reference). A signal in the aromatic region around 280 nm is consistent with the presence of Trp, Tyr, and Phe chromophores in an ordered environment, such as would be expected in the hydrophobic core of a folded protein. In general, assays for the affinity-purified expressed proteins that employ solely biophysical criteria have been designed based upon experience with the behavior of misfolded recombinant proteins.

It is preferable to further characterize the isolated domain by ^1H -NMR spectroscopy. Preferably, the isolated domain is in a moderately concentrated solution ($\sim 100 \mu\text{M}$). A high dispersion pattern of the proton resonance spectrum is reported to be characteristic of a well-folded polypeptide.

A time-course of amide hydrogen-deuterium exchange measurements can also be performed on the isolated domain. From this, it is possible to observe whether backbone NH groups are significantly protected within the domain. Significant protection is an indication that the hydrogen-bonded secondary structure is stabilized by tertiary interactions, which is consistent with a well-folded domain structure.

Finally, thermal denaturation experiments, monitored by intrinsic tryptophan fluorescence, can also be performed. These experiments are also capable of determining whether the isolated domain is a compact domain structure.

In principle, this is a general strategy. Thus, it can be used to parse many genes in the human genome that encode proteins of unknown biochemical function into their component domains and express correctly-folded polypeptide for structure/function studies. This general strategy can be easily modified to provide a high-throughput

5 method for validating candidate domains identified by the bioinformatics methods of the present invention. For a typical 10 - 30 kD protein domain, 500 or 600 MHz one-dimensional (1D) NMR spectra can be obtained in tens of minutes using only small
10 quantities (~ 200 µg) of protein. Using a continuous flow NMR probe with a
5 microcomputer-controlled chromatography pump and simple sample changer, it is possible to automatically screen 50 - 100 candidate domains per day for folded structure. Those candidate domains which exhibit chemical shift dispersion indicative
15 of ordered domain structure can then be further validated using the other biophysical techniques described above. An NMR spectrometer suitable for use in the present
10 invention is a Varian Unity 500 spectrometer.

20 C. High Level Expression And Isotopic Enrichment

Uniform biosynthetic enrichment with ^{15}N , ^{13}C and ^2H isotopes has been reported to be a prerequisite for the analysis of macromolecular structures by NMR spectroscopy. Some NMR strategies have also been reported to benefit from random
25 enrichment with ^2H isotopes. The principal obstacle for isotope-enriched protein production in most recombinant production systems is the high cost of the enriched media components (e.g. ^{13}C -glucose @ \$330/g), and the limiting possibilities for scale-up to controlled multi-liter fermenters. The less well-controlled conditions of shaker flask cultivations often result in lower protein production levels. The production of ^{15}N -
30 ^{13}C -, and/or ^2H -enriched proteins thus requires an efficient system cable of providing high level production of the desired protein in small-scale bioreactors.

Under one preferred embodiment, the present invention employs a bacterial production system for ^{15}N , ^{13}C -enriched recombinant proteins. Preferably, the bacterial production system is based on intracellular production of recombinant proteins in *E. coli*
35 as fusions to an IgG-binding domain analogue, Z, derived from staphylococcal Protein A (Nilsson *et al.*, *Protein Eng.* 1:107-113 (1987); Altman *et al.*, *Protein Eng.* 4:593-600 (1991), both of which are herein incorporated by reference). In this system,
40 transcription is initiated from the efficient promoter of the *E. coli trp* operon. This allows for efficient intracellular production of fusion proteins. These fusion proteins
30 can then be purified by IgG affinity chromatography. Using this approach it is possible to achieve high-level (40 - 200 mg/L) production in defined minimal media of a number of isotope-enriched proteins (see, for example, Jansson *et al.*, *J. Biomol. NMR* 7:131-141 (1996)).
45

5 Under another preferred embodiment, the recombinant isotope-enriched domain protein may be produced using pET plasmid expression vectors (Studier *et al.*, *J. Mol. Biol.* 189:113-130 (1986), herein incorporated by reference) under the control of the T7
10 RNA polymerase promoter (see, for example, Newkirk *et al.*, *Proc. Nat'l Acad. Sci. (U.S.A.)* 91:5114-5118 (1994); Chatterjee *et al.*, *J. Biochem.* 114:663-669 (1993); and Shimotakahara *et al.*, *Biochemistry* 36:6915-6929 (1997), all of which are herein incorporated by reference).

15 Under another preferred embodiment, ^{15}N , ^{13}C , ^2H -enriched recombinant proteins can be produced by acclimating a bacterial production system to grow in 95% $^2\text{H}_2\text{O}$. Recombinant bacterial production hosts [e.g., the BL21 (DE3) strain] can be
20 acclimated to grow in 95% $^2\text{H}_2\text{O}$ by successive passages in media containing increasing amounts of $^2\text{H}_2\text{O}$; protein production levels of acclimated bacteria grown in 95% $^2\text{H}_2\text{O}$ are identical to those obtained in H_2O . Using protiated [uniformly ^{13}C -enriched]-glucose as the carbon source, ^2H -enrichment levels of 70 - 80% can be achieved; high
25 incorporation of ^2H from the $^2\text{H}_2\text{O}$ solvent results from metabolic shuffling during amino acid biosynthesis. While the resulting proteins are not 100% perdeuterated, they are sufficiently enriched for the purpose of slowing ^{13}C transverse relaxation rates and enhancing the sensitivity for certain types of triple-resonance NMR experiments. 100% perdeuterated samples can also be produced using $^2\text{H}_2\text{O}$ solvent and [uniformly ^2H , ^{13}C -enriched]-glucose as the carbon source.

30 Under one preferred embodiment, such isotope enriched proteins can be renatured by the method of Kim *et al.* which employs *in situ* refolding of proteins immobilized on a solid support. Kim *et al.*, *Prot. Eng.* 10:445-462 (1997), herein incorporated by reference. The isotope enriched proteins can also be renatured by the
35 method of Maeda *et al.* which employs programmed reverse denaturant gradients. Maeda *et al.*, *Protein Eng.* 9:95-100 (1996); Maeda *et al.*, *Protein Eng.* 9:461-465 (1996), both of which are herein incorporated by reference. Under another preferred embodiment, the method of Kim *et al.* is coupled with the method of Maeda *et al.*
40 Under yet another preferred embodiment, "active" folding agents, such as the molecular chaperones GroEL/ES, dnaK, dnaJ, etc., may be used to assist in protein folding. Nilsson *et al.*, *Ann. Rev. Microbiol.* 45:607-635 (1991), herein incorporated by reference.

45 Preferably, the fusion vectors are constructed to interface with downstream refolding operations. Such vectors permit, for example, the binding of fusions to a solid
50 support even under harshly denaturing conditions, such as high concentrations of guanidine hydrochloride and dithiothreitol. For such purposes, the preferred class of

vector employs protein-RNA fusions. Such fusion proteins can be purified using oligonucleotide affinity columns with high specificity in the presence of chaotropic agents and strongly reducing conditions.

Under another preferred embodiment, other, non-bacterial, microbial systems, e.g., *Pichia*-based expression systems are employed. Kocken *et al.*, *Anal. Biochem.* 239:111-112 (1996); Munshi *et al.*, *Protein Expr. Purif.* 11:104-110 (1997); Laroche *et al.*, *Bio/Technology* 12:1119-1124 (1994) Cregg *et al.*, *Bio/Technology* 11:905-910 (1993), all of which are herein incorporated by reference.

Once the protein domain of interest has been expressed at high levels, it is necessary to purify large quantities of the protein domain for subsequent characterization. Preferably, at least 5-10 mg of the protein domain of interests is purified. More preferably, at least 50 mg of the protein domain of interest is purified.

Methods for preparing large quantities of a given protein of sufficient purity for domain structure modeling are generally known to those of skill in the art. Although not all methods for protein purification are applicable to a given protein of interest, it is generally understood that the following methods represent preferred embodiments: affinity chromatography, ammonium sulfate precipitation, dialysis, FPLC chromatography, ion exchange chromatography, ultracentrifugation, etc. For a general review of protein purification methodologies, see Burgess, *Protein Purification*, In: Oxender *et al.* (Eds.), *Protein Engineering*, pp. 71-82, Liss (1987); Jakoby, (Ed.), *Methods Enzymol.* 104:Part C (1984); Scopes, *Protein Purification: Principles and practice* (2nd ed.), Springer-Verlag (1987), and related texts, all of which are herein incorporated by reference.

D. Rapid Screening Of NMR And Crystallization Properties

One common problem for both NMR analysis and crystallization is poor solubility and/or slow precipitation of the protein sample. These properties are highly dependent on the pH, ionic strength, reducing agent concentration, and other properties of the buffer solvent. Thus, it is preferable to optimize these conditions to maximize solubility for NMR analysis and to optimize the conditions for protein crystallization.

Under one of the preferred embodiments of the present invention, the optimization experiments are conducted with an array of microdialysis buttons to rapidly scan a plurality of standardized buffer conditions to identify those most suitable for NMR studies and/or crystallization of each domain construct (Bagby, *J. Biomol. NMR* 10:279-282 (1997), incorporated by reference in its entirety). Preferably, each

microdialysis button contains at least 1 μ L of a ~1 mM protein solution. More preferably, each microdialysis button contains at least 5 μ L of a ~1 mM protein solution. The microdialysis buttons of the present invention are commercially available. Preferably, each microdialysis button is dialyzed against about 50 ml of dialysis buffer, such as in a 50 ml conical tube (Falcon). Preferably, the dialysis is performed at 4°C. However, the dialysis can be performed at temperatures ranging from 4°-40°C. Because NMR studies are routinely performed at room temperature for extended lengths of time, it is preferable that the protein remain in solution under these conditions.

Preferably, the protein samples are initially prepared in buffers containing 50% glycerol (which is not suitable for NMR studies but generally provides good solubility) and then dialyzed against different buffers containing little or no glycerol. With respect to NMR and X-ray crystallography studies, it is understood that a person of skill in the art would know what buffers could be used to prepare the protein for study. The skilled artisan typically has a set of 50-100 standard buffers which are used to prepare protein samples for subsequent studies. These buffers can then be modified if necessary to optimize the protein preparation. The ability of a given protein to remain soluble at high concentration or form suitable crystals is dependent on the pH of the solution, as well as the concentration of different salts, buffers, reagents, and temperature. Thus, the "button test" represents a preferred embodiment because it facilitates the rapid screening of a multitude of conditions.

This "button test" analysis typically requires 5 - 10 mg of protein sample and can be completed in a few days. Preferably, multiple samples are analyzed in parallel. Preferably, the protein samples are analyzed under a dissecting microscope to determine whether the protein has remained in solution or whether the protein has aggregated. Using the "button test" of the present invention, a single technician could score solubility properties in 100 different buffers for ~20 domains per week. Under the another preferred embodiment, these screens can be carried out using state of the art laboratory automation technology.

Alternatively, the protein domain of interest is lyophilized and then resuspended in an appropriate buffer.

Having identified the conditions under which the protein domain of interest is soluble, dynamic light scattering can be used to examine its dispersive properties and aggregation tendency in different buffer conditions. Ferré-D'Amaré *et al.*, *Structure* 15:357-359 (1994), herein incorporated by reference. Alternatively, Trp or Tyr fluorescence anisotropy can be used to measure rotational diffusion which is another measure of aggregation.

5 The "domain trapping" approach of the present invention includes an evaluation of NMR properties, and all of the protein samples which pass this stage of the process will already meet basic spectroscopic quality criteria. Standard criteria used to
10 determine the basic spectroscopic quality of a given protein, which are known to those of skill in the art, include a good dispersion pattern and a narrow peak width, etc.

5 Preferably, gel filtration chromatography and dynamic light scattering data are collected during the course of domain purification. Such data provide information about the oligomerization state of the domain being studied.

15 For domains of the appropriate size (< ~30 kD), isotopically enriched samples are scored in terms of their suitability for structure determination by NMR using standard 2D HSQC, 2D NOESY, and/or 2D CBCANH triple-resonance spectra. The
20 protein samples that provide good quality data for these NMR experiments are expected to provide good data in the full set of experiments required for automated structure determination. For each ^{15}N , ^{13}C enriched domain, this evaluation typically requires at
15 least 5 - 10 mg of sample, and approximately 6 hours of NMR data collection. Preferably, the evaluation is performed on about 10 mg of sample. Thus, ~20 domains can be evaluated per "spectrometer-week" using the methods of the present invention.
25 A "spectrometer-week", as used herein, means one skilled technician, working on one NMR machine would be able to evaluate approximately 20 domains in a given week.

30 Preferably, domains for structure determination by NMR are selected in an opportunistic manner, prioritizing those that provide high quality NMR data in the screens outlined above. Although some of the constructs that are generated may not be amenable to rapid structural analysis, it has been estimated that well over 50% of
35 domains that are "trapped" by the process outlined above exhibit properties suitable for NMR or X-ray analysis. As these domains are derived from specific target genes associated with human diseases (discussed below) the chances of obtaining important new protein structures by this process are very high. Domains that provide diffraction
40 quality crystals and which are not amenable to rapid analysis by NMR can be analyzed by X-ray crystallography.

30 **E. Computer Software And Related NMR Technologies
For Fully Automated Analysis Of Protein Structures
From NMR Data**

45 The present invention employs advanced NMR data collection and automated analysis technologies. These data collection and automated analysis technologies
35 greatly accelerate the process of protein structure determination. Included within these

technologies is a family of easy to use pulsed-field gradient triple resonance NMR experiments for rapid analysis of protein resonance assignments. See, for example, Montelione *et al.*, *Proc. Natl. Acad. Sci. (U.S.A.)* 86:1519-1523 (1989); Montelione *et al.*, *Biopolymers* 32:327-334 (1992); Montelione *et al.*, *Biochemistry* 31:236-249 (1992); Lyons *et al.*, *Biochemistry* 32:7839-7845 (1993); Rios *et al.*, *J. Biomol. NMR* 8:345-350 (1996); Tashiro *et al.*, *J. Mol. Biol.* 272:573-590 (1997); Shimotakahara *et al.*, *Biochem.* 36:6915-6929 (1997); Laity *et al.*, *Biochem.* 36:12683-12699 (1997); Feng *et al.*, *Biochem.* 37:10881-10896 (1998); and Swapana *et al.*, *J. Biomol. NMR* 9:105-111 (1997), all of which are herein incorporated by reference. These data collection and automated analysis technologies further include a fully automated strategy for determining NMR resonance assignments in proteins. Zimmerman *et al.*, *Curr. Opin. Struct. Bio.* 5:664-673 (1995); and Zimmerman *et al.*, *J. Mol. Biol.* 269:592-610 (1997), both of which are herein incorporated by reference.

Preferably, the data collection and automated analysis technologies of the present invention employ multiple-quantum coherences in triple resonance for enhanced sensitivity. Swapna *et al.*, *J. Biomol. NMR* 9:105-111 (1997); Shang *et al.*, *J. Amer. Chem. Soc.* 119:9274-9278 (1997), both of which are herein incorporated by reference.

1. AUTOASSIGN: Artificial Intelligence Methods For Automated Analysis Of Protein Resonance Assignments

Resonance assignments form the basis for analysis of protein structure and dynamics by NMR (Wüthrich, K., *NMR of Proteins and Nucleic Acids*, John Wiley & Sons, New York, New York (1986), herein incorporated by reference) and their determination represents a primary bottleneck in protein solution structure analysis. However, the introduction of multi-dimensional triple-resonance NMR has dramatically improved the speed and reliability of the protein assignment process. Montelione *et al.*, *J. Magn. Res.* 83:183-188 (1990); Ikura *et al.*, *Biochem. Pharmacol.* 40:153-160 (1990); Ikura *et al.*, *FEBS Letters* 266:155-158 (1990); Ikura *et al.*, *Biochem.* 29:4659-4667 (1990); Tashiro *et al.*, *J. Mol. Biol.* 272:573-590 (1997); Shimotakahara *et al.*, *Biochem.* 36:6915-6929 (1997); Laity *et al.*, *Biochem.* 36:12683-12699 (1997); Feng *et al.*, *Biochem.* 37:10881-10896 (1998), all of which are herein incorporated by reference.

Preferably, the present invention employs AUTOASSIGN, an expert system that determines protein ¹⁵N, ¹³C, and ¹H resonance assignments from a set of three-dimensional NMR spectra. Zimmerman *et al.*, *Proceedings of the First International Conference of Intelligent Systems for Molecular Biology* 1:447-455 (1993); Zimmerman

5 *et al.*, *J. Biomol NMR* 4:241-256 (1994); Zimmerman *et al.*, *Curr. Opin. Struct. Bio.*
5:664-673 (1995); Zimmerman *et al.*, *J. Mol. Biol.* 269:592-610 (1997), all of which are
10 herein incorporated by reference. AUTOASSIGN has been copyrighted by Rutgers, the
State University of New Jersey. Alternatively, the present invention can employ one of
5 the following expert systems for the automated determination of protein ^{15}N , ^{13}C , and
 ^1H resonance assignments from a set of three-dimensional NMR spectra. These include
a modified version of FELIX which is available from Molecular Simulation (San Diego,
15 CA) (Friedrichs *et al.*, *J. Biomol. NMR* 4:703-726 (1994), incorporated by reference in
its entirety). CONTRAST which is available from the world wide web at
10 <<www.bmrb.wisc.edu/macros/soft_contrast.html>> (Olsen and Markley, *J. Biomol.*
NMR 4:385-410 (1994), incorporated by reference in its entirety), and a series of small
programs described by Meadows, *J. Biomol. NMR* 4:79-86 (1994), incorporated by
20 reference in its entirety.

AUTOASSIGN is implemented in the Allegro Common Lisp Object System
15 (CLOS) and requires a lisp compiler (available from Franz, Inc.) for execution. The
software utilizes many of the analytical processes employed by NMR spectroscopists,
25 including constraint-based reasoning and domain-specific knowledge-based methods.
Fox *et al.*, *The Sixth Canadian Proceedings in Artificial Intelligence* 1986; Nadel *et al.*,
Technical Report, DCS-TR-170, Computer Science Department, Rutgers Univ. (1986);
20 Kumar *et al.*, *Artificial Intelligence Mag.*, Spring, 32-44 (1992), all of which are
incorporated by reference in their entirety.

Input to AUTOASSIGN includes a peak-picked 2D (H-N)-HSQC spectrum and
the following seven peak-picked 3D spectra: HNCO, CANH, CA(CO)NH, CBCANH,
35 CBCA(CO)NH, H(CA)NH, and H(CA)(CO)NH. This family of triple-resonance
25 experiments can be used together with AUTOASSIGN to automatically determine
extensive sequence-specific ^1H , ^{15}N , and ^{13}C resonance assignments for several proteins
ranging in size from 8 kD to 17 kD. Zimmerman *et al.*, *J. Mol. Biol.* 269:592-610
(1997); Tashiro *et al.*, *J. Mol. Biol.* 272:573-590 (1997); Shimotakahara *et al.*, *Biochem.*
40 36:6915-6929 (1997); Laity *et al.*, *Biochem.* 36:12683-12699 (1997); Feng *et al.*,
30 *Biochem.* 37:10881-10896 (1998). The program handles some of the very challenging
problems encountered in automated analysis, including missing spin systems, spin
systems that overlap even in the 3D spectra, and extra spin systems due to multiple
45 conformations of the folded protein structure (e.g. X-Pro peptide bond cis/trans
isomerization). Execution times on a Sun Sparc 10 workstation range from 16 to 360
35 sec, depending on the complexity of the problem analyzed by the program. Preferably,
the NMR spectrometer of the present invention is equipped with three channels and a

fourth frequency synthesizer for carbonyl decoupling. Under another preferred embodiment, the NMR spectrometer of the present invention is equipped with four channels.

In the present invention, the AUTOASSIGN program provides for automated analysis of resonance assignments for atoms of the polypeptide backbone. Preferably, the AUTOASSIGN program of the present invention provides for fully automated analysis of resonance assignments. Having established assignments for the backbone atoms of each amino acid in the protein sequence, it is relatively straightforward to extend from these to sidechain ^1H and ^{13}C resonance assignments using 3D HCCH COSY, HCCH-TOCSY, and HCC(CO)NH-TOCSY NMR experiments. Preferably, the AUTOASSIGN program of the present invention handles automated analysis of these sidechain resonance assignments. It is additionally preferred that 3D ^{15}N -edited NOESY and 3D ^{13}C -edited NOESY data are collected and automatically analyzed to confirm the resonance assignments.

Under one of the preferred embodiments of the present invention, AUTOASSIGN is designed to implement strategies that allow complete resonance assignments to be obtained with fewer NMR spectra. For example, sensitivity enhanced versions of HCCNH-TOCSY and HCC(CO)NH-TOCSY experiments can provide the complete set of information required for the determination of resonance assignments. This reduces the total data collection time required for determining backbone resonance assignments from the current 7 - 10 days to about half of this time. Zimmerman *et al.*, *J. Biomol. NMR* 4:241-256 (1994); Lyons *et al.*, *Biochemistry* 32:7839-7845 (1993), both of which are herein incorporated by reference.

Perdeuteration greatly lengthens the ^{13}C transverse relaxation rates, allowing for higher sensitivity in these triple-resonance experiments. Grzesiek *et al.*, *J. Biomol. NMR* 3:487-493 (1993); Yamazaki *et al.*, *Eur. J. Biochem.* 219:707-712 (1994), both of which are herein incorporated by reference. It has been demonstrated that significant sensitivity-enhancement (2 - 5 fold) can be obtained with triple-resonance experiments by perdeuteration of the protein samples. Preferably, the automated assignment strategy, described herein, will utilize ^2H , ^{13}C , ^{15}N -enriched proteins prepared with protiated ^{15}N -H amide groups, together with deuterium-decoupled triple resonance NMR experiments. Under one embodiment, the amide NH group in the perdeuterated protein exchanges rapidly with the solvent H_2O used in the course of the protein purification to yield the protiated ^{15}N -H amide groups. This strategy can provide completely automated analysis of resonance assignments for the carbon and nitrogen skeleton of the protein. Having determined these assignments, analysis of resonance

5 assignments for the attached hydrogen atoms can be completed using HCCH-COSY,
10 HCCH-NOESY, and HCCH-TOCSY experiments. Correction factors for ^2H -isotope
shift effects for each carbon site of the 20 amino acids can be determined using data
15 from model proteins. Preferably, the complete carbon resonance assignments in their
20 protiated forms have already been determined for these model proteins.

Preferably, the present invention utilizes high temperature superconducting
probes. First generation versions of these probes are currently being marketed by
15 Varian NMR Inst. Inc. and Bruker Inst. Such probes in combination with the above-
described technological advances reduce the time required for determining complete
20 backbone and sidechain H, C, and N assignments to less than one week per domain.

2. Software For Automated Analysis Of Protein Structures From NMR Data

25 Having completed the resonance assignments for a particular protein, the next
step of the structure determination process of the present invention involves analyzing
secondary structure (i.e. α -helices, β -sheets, turns, etc.). The chemical shifts themselves
25 are often sufficient to allow identification of these features of secondary structure in the
protein. Spera, *J. Amer. Chem. Soc.* 113:5490-5492 (1991); Wishart *et al.*, *J. Biomol.*
NMR 6:135-140 (1995), both of which are herein incorporated by reference. This
30 information can be combined with other bioinformatics data derived from the protein
sequence to narrow the number of possible mappings of the protein to known chain
folds, and possibly even to identify the protein's biochemical function.

The principal sources of information used for the structure determination of
protein domains are nuclear Overhauser effect (NOE) data arising from magnetic
35 dipole-dipole interactions between hydrogen atoms in the structure of the protein.

25 Interpretation of these data from multidimensional NOE spectroscopy (NOESY) spectra
requires the resonance assignments, which will be obtained (as described above) in an
automated manner. Preferably, the present invention employs software for automated
analysis of NOESY spectra and the generation of input files for rapid structure
40 calculations using stimulated annealing of experimental constraint functions with
30 molecular dynamics calculations.

The problems encountered in automatically analyzing NOESY spectra are due
largely to spectral overlaps, i.e., it is often the case that several hydrogen atoms have
45 very similar resonance frequencies. One of the preferred approaches to resolving this
problem is to use 3D (or 4D) ^{15}N - or ^{13}C -resolved NOESY experiments (Clare *et al.*,
35 *Ann. Rev. Biophys. Biophys. Chem.* 20:29-63 (1991); Clare *et al.*, *Prog. Biophys. Mol.*

Bio. 62:153-184 (1994); Clore *et al.*, *Methods Enzymol.* 239:349-363 (1994), all of which are herein incorporated by reference), in which one (or both) of the two protons involved in the NOE interaction is resolved in a third (or fourth) frequency dimension based on the frequency of the ¹⁵N or ¹³C nucleus to which it is covalently bound.

5 Symmetry features of the 3D ¹³C-edited spectra can also be used to great advantage.

Another preferred approach to resolving ambiguities that arise in assigning NOESY cross peaks to specific pairs of interacting hydrogen atoms is to use the secondary structure (i.e. α helix, β strand, etc.) to predict NOEs that are expected and to use these structural predictions to guide the analysis of NOESY spectra. Meadows *et al.*, *J. Biomol. NMR* 4:79-96 (1994), herein incorporated by reference.

A third preferred approach is to use a low-resolution structure of the protein obtained in a first pass analysis of the uniquely assigned NOESY cross peaks to identify candidate assignments of the remaining unassigned NOESY cross peaks which are inconsistent with the low-resolution structure.

15 The approaches outlined above are those that are routinely used by a human expert in the analysis of NOESY spectra. Under the preferred embodiment, the reasoning processes of those approaches are encoded into the software of the present invention. Preferably, the software program of the present invention is a C++ program. AUTO_STRUCTURE is a C++ program that analyzes 2D and 3D NOESY spectra to identify unique NOESY crosspeak assignments (Gaetano Montelione, Y. Huang and Robert Tejero (Rutgers, The State University of New Jersey)). The program then uses these crosspeak assignments to create distance-constraint input files for simulated annealing structure calculations. AUTO_STRUCTURE can also use a low-resolution (or homology-modeled) structure of the protein to filter the list of NOESY crosspeaks that are not uniquely assigned, removing potential NOE assignments that are severely inconsistent with the low-resolution structure. AUTO_STRUCTURE propagates the structural constraints imposed by the uniquely assigned NOEs to determine assignments of otherwise ambiguous NOEs. AUTO_STRUCTURE can successfully analyze NOESY spectra and, in an iterative fashion, automatically generate 3D structures of simple polypeptides. Other auto structure programs for NOESY analysis that can be used in the present invention include GARANT (Wuthrich (ETH, Zurich, Germany), incorporated by reference in its entirety), ARIA (Michael Nilges, *J. Mol. Biol.* 245:645-660 (1995), incorporated by reference in its entirety) and NOAH (Mumenthaler and Braun, *J. Mol. Bio.* 254:465-420 (1995), incorporated by reference in its entirety).

35 Preferably, the auto structure program of the present invention provides for automated analysis of protein or protein domain structures. Under a more preferred

embodiment, the auto structure program of the present invention further contains sophisticated reasoning processes which can assist in resolving ambiguous NOESY crosspeak assignments in the absence of even a low resolution 3D structure. Preferably, this includes (i) the propagation of structural constraint information inherent in the secondary structure analysis stemming from the resonance assignments and (ii) the application of pattern recognition algorithms.

F. Mapping New Domain Structures To Proteins In The Protein Data Base (PDB) With Similar Structures And Biochemical Functions

Preferably, the resulting domain structures derived from NMR or X-ray crystallographic analyses are compared with the PDB or other suitable databases of known protein structures using an algorithm for 3D-structure homology matching. Examples of publicly available PDBs suitable for use in the present invention include the Protein Data Base (PDB), which can be found at <http://www.pdb.bnl.gov/>. Algorithms for 3D-structure homology matching suitable for use in the present invention include the DALI analysis program (Holm *et al.*, *J. Mol. Biol.* 233:123-138 (1993), herein incorporated by reference), the CATH analysis program (Orengo, C. A., *Structure* 5:1093-1108 (1997), herein incorporated by reference), VAST (<http://www.ncbi.nlm.nih.gov/Structure/vast.html>; Gibrat *et al.*, *Current Opinion in Structural Biology* 6: 377-385 (1996); and Madej *et al.*, *Proteins* 23: 356-369 (1995), all of which are incorporated by reference in their entirety) or similar algorithms for 3D-structure homology matching.

DALI compares "contact maps" of protein structures to identify homologies in 3D structure and provides a list of PDB entries with high match scores. Based on current "hit" rates by newly-determined structures against already known folds (Holm *et al.*, *Methods Enzymol.* 266:653-662 (1996); Holm *et al.*, *Science* 273:595-603 (1996), both of which are herein incorporated by reference), it is expected that greater than 50% of the structures will show significant structural and functional homology to proteins of known structure and function.

In order to facilitate and enhance the ability to identify common biochemical functions for these DALI hits, it is preferable to develop a structure-function knowledge base (Figure 1), correlating each protein structure in the PDB with the set of biochemical functions that have been associated with that protein in the published scientific literature. Where information is available, this knowledge base will also correlate the portions of these known protein structures with corresponding specific

5 biochemical functions (e.g., enzymatic active sites or nucleic-acid binding loops). This
fold-function knowledge base is applicable to a wide range of structural bioinformatics
10 applications, and of significant utility to the nascent industry of structural
bioinformatics.

5 Once novel protein domains with clear homologies to better-characterized
counterparts have been identified, the proposed functions can be validated using
15 biochemical assays. For example, if a protein looks like a member of the galactosyl
transferase family, the protein will be tested for radioactive UDP-galactose (or other
carbohydrate) binding, if it looks like a lipase, the protein will be tested for lipid binding
10 and/or hydrolysis activity, and so on.

20 **G. Integration Into A Large-Scale, High-Throughput
"Engine" For Structural And Functional Analysis Of
Hundreds Of Human Genes**

15 Under one preferred embodiment, the present invention provides for a "structure
- function analysis engine" capable of high-throughput discovery of biochemical
25 functions of new human disease genes and genes of unknown function.

Using conventional methodology, the skilled artisan may be able to determine
the 3D structure of one protein per year. However, using the methodology of the
present invention, it is possible to determine the 3D structure of far greater than one
30 protein per year. Under optimal conditions, the present invention will enable a properly
equipped laboratory to generate the 3D structure of one protein per month per NMR
machine. As used herein, "high-throughput" refers to the ability to determine the 3D
structures of protein domains of unknown function at a rate which is faster than the rate
35 at which a skilled artisan could determine a protein structure using traditional
methodologies.

One of the central features of the present invention is that it is highly scaleable.
Under one of the preferred embodiments, the high-throughput "engine" consists of a
40 dedicated laboratory staffed with artisans skilled in relevant arts (e.g., NMR and X-Ray
crystallography, molecular biology, biochemistry, etc.). Preferably, such a laboratory is
30 further equipped with state of the art equipment for the sequencing, sub-cloning,
expression, purification, screening and analysis of the protein domains of interest. The
rate limiting component of this high-throughput "engine" is the number of NMR
45 machines within the laboratory. Thus, the rate at which protein domains can be
characterized will increase with the addition of additional NMR machines. Unlike
35 conventional methodology, the present invention provides a method for determining the

5 3D structure of unknown protein domains whose rate is not solely dependent on the number of artisans skilled in 3D protein structure determination.

10 The rate of domain characterization increases as each of the tasks which are presently conducted by hand are automated. For example, under one of the preferred
5 embodiments, the parsing of the unknown gene into its component domains is facilitated through the use of advanced sequence analysis algorithms. Under another of the preferred embodiments, the rate of domain characterization is increased through the use of improved computer software for the automated analysis of NMR datapoints.

15 Although the present invention is drawn to using NMR to determine protein
10 structure and function, it is to be understood that a person of skill in the art could perform similar analysis using X-ray crystallography to practice the present invention. Shapiro and Lima, *J. Structure* 6:265-267 (1998); Gaasterland, *Nature Biotech.* 16:625-
20 627 (1998); Terwilliger *et al. Prot. Sci.* 7:1851-1856 (1998); Kim, *Nature Structure Biology (Synchrotron Supp.)*: 643-645 (1998), all of which are incorporated by
15 reference in their entirety.

25 III. SPECIFIC GENE TARGETS

Preferably, the specific gene targets that will be analyzed using the present invention will be genes that are known to be involved in human diseases but for which the biochemical function and three-dimensional structures of the proteins encoded by
30 20 the genes are not available. These protein domains will be analyzed using the high-throughput "structure - function analysis engine" of the present invention. The resulting structural and functional information will be critical in developing pharmaceuticals targeted to these human gene products.

35 Although the present invention is principally drawn to human genomic, cDNA
25 and mRNA sequences, it is to be understood that the present invention is generically applicable to genomic, cDNA and mRNA sequences of any living organism or virus.

40 Although the present invention is capable of determining the function of any given protein or protein domain, the preferred biomedical gene targets of the present invention include Alzheimer's β peptide precursor protein (APP). Additional preferred
30 biomedical gene targets include but are not limited to those genes implicated in neoplastic, neurodegenerative, metabolic, cardiovascular, psychiatric and inflammatory disorders. The genomes/genes of infectious agents, such as pathogenic microbes,
45 pathogenic fungi and pathogenic viruses, are also preferred targets for study.

By focusing on medically important diseases, it is anticipated that the present invention will greatly facilitate the identification of protein targets for subsequent drug discovery efforts.

Having now generally described the invention, the same will be more readily understood through reference to the following examples which are provided by way of illustration and are not intended to be limiting on the present invention.

EXAMPLE 1

PARSING OF THE APP GENE INTO DOMAIN-ENCODING REGIONS

A. Parsing By The Exon Phase Rule

The human amyloid beta peptide precursor (APP) protein gene (Yoshikai *et al.*, *Gene* 87:257-263(1990)) was subjected to a parsing analysis with respect to the phases of its exon-exon boundaries:

<u>Exon-exon boundary</u>	<u>Phase</u>
1 - 2	0
2 - 3	0
3 - 4	1
4 - 5	0
5 - 6	2
6 - 7	1
7 - 8	1
8 - 9	1
9 - 10	0
10 - 11	0
11 - 12	0
12 - 13	0
13 - 14	1
14 - 15	1
15 - 16	1
16 - 17	0
17 - 18	0

Using the exon phase rule, only exons or exon combinations that start or stop in the same phase are allowed. For example, exon 7 or exons 7+8 are potential domain

5 encoding regions with phase 1 boundaries. Likewise, exon 10, exons 10+11, and exons 10+11+12 would be potential domain encoding regions with phase 0 boundaries.

10 **B. Exon Phase And The Alternative Splicing Rule**

5 The APP gene is reported to be alternatively spliced. The longest polypeptide encoded by the APP gene is 770 amino acids long, and shorter isoforms exist that are missing the amino acids encoded by exons 7, 8, and/or 15 (Sandbrink *et al.*, *Ann. NY Acad. Sci.* 777:281-287 (1996), herein incorporated by reference). All of these exons 15 which are alternatively spliced are bounded by phase 1 termini. Alternative splicing must be done in such a way as to not disrupt the integrity of the holoprotein (i.e., 10 without destroying essential folding information). The fact that all alternatively spliced exons have phase 1 termini implies that domain boundaries may be congruent with 20 phase 1 exon boundaries, that is, phase 1 exon boundaries in this particular gene are candidate boundaries of domain encoding regions.

25 **C. Setting The Phase With Known Internal Domain Structures**

15 Exon 7 of APP is known to encode a complete domain for a Kunitz-type serine protease inhibitor (Hynes *et al.*, *Biochemistry* 29:10018-10022 (1990)). The Kunitz 25 inhibitor is a domain that has been combinatorially shuffled around in various genes during evolution (Patty, L. *Curr. Opin. Struct. Biol.* 1:351-361 (1991)), and for the reasons given above it would have to be inserted only into proteins with other domains 30 of the same phase in order to not disrupt gene expression. Therefore, this analysis is also consistent with APP being composed of domains which are bounded by phase 1 exon termini.

35 **D. The "N-Terminus First" Strategy Of Parsing**

35 In order to reduce the combinatorial complexity of the parsing problems, an "N-terminus first" strategy is preferred. In this parsing strategy, expression constructs of 25 putative domains are made starting from the N-terminus of the protein and extending to the likely C-termini as predicted by the above rules. These constructs are put through 40 the "domain trapping" test of the present invention in order to identify the first N-terminal domain. Then, once the first N-terminal domain is identified, a second set of 30 constructs commencing from the C-terminus of the first N-terminal domain is made, and so on.

45 In the case of APP, the N-terminus of the protein starts with exon 2 because exon 1 encodes a signal peptide. Therefore, the possible domain constructs that ended in phase 1 boundaries were exons 2-3 and exons 2-6 (exon 7 was known to encode the 50

Kunitz inhibitor domain). By the domain trapping criteria exons 2-3 were found to encode the first N-terminal domain, so a second construct composed of exons 4-6 was made and found to contain the second domain of APP, and so on. A summary of the APP domains identified by this combination of parsing and domain trapping is given below:

<u>Domain</u>	<u>Encoding Exons</u>
1 (N-terminal domain)	2-3
2	4-6
3 (Kunitz inhibitor)	7
4	8
etc.	

EXAMPLE 2

EXPRESSION AND PURIFICATION OF AN ISOLATED DOMAIN

The putative domain regions identified in Example 1 are sub-cloned into the secretion-based protein A fusion expression system and purified. Nilsson *et al.*, *Methods Enzymol.* 185:144-161 (1990), herein incorporated by reference.

EXAMPLE 3

EXPRESSION AND PURIFICATION OF AN ISOLATED DOMAIN FOR NMR ANALYSIS

A. Protein Expression

E. coli strain RV308 is used as the bacterial expression host. Competent RV308 cells are transformed with pHAZY plasmid containing the NTD 2-3, Z domain insert. Cells are grown overnight at 37°C on LB agar plates supplemented with 100 µg/ml ampicillin (Sigma). Fresh transformants are used to inoculate seed cultures in 2 x TY media (16 g/l typtone, 10 g/l yeast extract, and 5/g NaCl) supplemented with 100 µg/ml ampicillin. Cultures are grown overnight at 30°C in 250 ml baffled flasks. A ratio of 1 to 25 is used to inoculate expression cultures. For 1 liter of MJ media expression culture (2.5 g/l ¹⁵NH₄ sulfate (>98% purity), 0.5 g/l sodium citrate, 100 mM potassium phosphate buffer, pH 6.6, supplemented with 5 g/l ¹³C-glucose (>98% purity), 1 g/l magnesium sulfate, 70mg/l thiamine, 1 ml of 1000 x trace elements solution, 1 ml of 1000 x vitamin solution, and 100 mg/l ampicillin), 40 ml of seed culture is spun down by centrifugation. Bacterial pellets are washed, resuspended in fresh MJ media, and used to inoculate expression cultures. Cultures are grown at 30° in

2 l baffled flasks and induced at OD⁵⁵ 0.9 – 1.0 with indole acrylic acid to a final concentration of 20 mg/l. Cultures are harvested 15 hours after induction by centrifugation. Bacterial pellets are stored at 20°C until purification.

B. Protein Purification

Bacterial cells are resuspended in 100 ml of 25 mM Tris, pH 8.0, 5 mM EDTA, 0.5% Triton X-100 and sonicated continuously for 9 minutes. Released inclusion bodies are pelleted by centrifugation and washed with fresh sonication buffer. Inclusion bodies were then solubilized with 7 M guanidine HCl and 10 mM DTT. Centrifugation is used to pellet any undissolved material. Guanidine and DTT are then diluted twenty fold by dialysis against twenty volumes of 10 mM HCl.

IgG affinity purification is used to purify the NTD 2-3, Z domain fusion from any contaminating proteins. The 10 mM HCl protein solution is neutralized to > pH 7 with 1 M Tris, pH 8.0. The sample is then applied to an IgG sepharose column (Pharmacia) pre-equilibrated with TST buffer. The column is washed with 10 bed volumes of TST (50 mM Tris, 150 mM NaCl, and 0.05% TWEEN™ 20) followed by 2 bed volumes of 5 mM ammonium acetate, pH 5.0. Finally, the protein is eluted with 0.5 M acetic acid, pH 3.4. In preparation for refolding, the protein eluate is neutralized to pH 8.0 with solid Tris, and an equal volume of 7 M guanidine is added to bring the final guanidine concentration to 3.5 M.

Refolding of the protein is carried out by using dialysis to slowly dilute out the guanidine HCl while slowly introducing the refolding buffer. Firstly, Spectra/POR dialysis tubing with a MWCO of 6000-8000 is soaked overnight in water in order to remove glycerol. Next, the protein solution is loaded into the primed tubing and dialyzed against fresh refolding buffer. The dialysis reaction is incubated for two days at 4°C with magnetic stirring. Refolded protein is then concentrated using an IgG sepharose column pre-equilibrated with TST buffer. Bound protein is eluted with 0.5 M acetic acid and collected in fractions in order to keep the volume as low as possible. Refolded fusion protein is then further purified by gel filtration on a Pharmacia Superdex 75 FPLC column using 300 mM ammonium bicarbonate, 0.1 mM copper sulfate as the buffer. Fractions corresponding to the fusion protein are pooled, and the protein is quantitated using the optical density at 280 nm.

Cleavage of the fusion protein is carried out using Genenase I (NEB), a variant of subtilisin BPN'. Fusion protein is buffer exchanged into Genenase buffer, 20 mM Tris, pH 8.0, 200 mM NaCl, 0.02% NaN₃, using an Amicon stir cell. The protein concentration is adjusted to 2 mg/ml and Genenase is added to a concentration of 0.2

5 mg/ml. The reaction is incubated at room temperature for 4 days and the extent of
cleavage was followed using SDS-PAGE. Cleaved NTD 2-3 is separated from
uncleaved fusion and Z domain by passing the solution over an IgG column and
10 collecting the unbound NTD 2-3 in the flow through. The NTD is then purified from
5 Genenase by gel filtration on a Pharmacia Superdex 75 FPLC column using 300 mM
ammonium bicarbonate, 0.1 mM copper sulfate as the buffer.

EXAMPLE 4

DOMAIN TRAPPING: CHARACTERIZATION OF AN ISOLATED DOMAIN

10 Characterization of an isolated domain (NTD2-3) from the Alzheimer's amyloid
precursor protein (APP) by circular dichroism measurements in the far UV shows an
20 ellipticity minimum at 222 nm, indicative of α -helical secondary structure (Figure 2A).
Of even greater significance, CD measurements at longer wavelengths reveal a clear
signal in the aromatic region around 280 nm, consistent with the presence of Trp, Tyr,
15 and Phe chromophores in an ordered environment such as would be expected in the
hydrophobic core of a folded protein (Figure 2B). A moderately concentrated solution
25 (~100 μ M) of the isolated N-terminal domain is further characterized by one-
dimensional 1 H-NMR. The isolated recombinant APP N-terminal domain exhibits high
dispersion of the proton resonances, which is a signature of well-folded polypeptides
30 (Figure 3).

A time-course of amide hydrogen-deuterium exchange measurements is
performed. From this, it is observed that many backbone NH groups exhibit significant
protection, indicating hydrogen-bonded secondary structure stabilized by tertiary
35 interactions consistent with a well-folded domain structure (Figure 4). Finally, thermal
denaturation experiments, monitored by intrinsic tryptophan fluorescence, are
25 performed. These experiments show that the recombinant APP NTD2-3 domain
undergoes a cooperative thermal unfolding transition, with a T_m of approximately 60° C,
indicative of a compact domain structure (Figure 5).

40 Thus, using biophysical data alone, it is demonstrated that the NTD2-3 domain
of APP, encoded by exons 2 and 3, is expressed as a well ordered tertiary structure.
30 Chiang *et al.*, *Neurobiol. Aging*, Supplement Vol. 17, No. 4S, abstract 393 (1996).
Similar studies indicate that the next APP N-terminal domain is encoded by exons 4-6,
45 the third (Kunitz) domain by exon 7, and so on.

EXAMPLE 5**NMR CHARACTERIZATION OF THE NTD 2-3 DOMAIN**

For NMR studies NTD 2-3 is concentrated to concentrations greater than 10 mg/ml. Gel filtration pure NTD 2-3 is first buffer exchanged into a NMR compatible buffer, 20 mM potassium phosphate, pH 6.5 using an Amicon stir cell. The protein solution is then concentrated to an appropriate volume based on the amount of protein present using the Amicon 50 and Amicon 3 stir cells. The final protein concentration is confirmed by optical density at 280 nm.

NMR ^{15}N -HSQC spectra is collected on a Varian Unity 500 spectrometer. The ^{15}N -HSQC spectral analysis is shown in Figure 6. The good dispersion in both the ^{15}N and ^1H dimensions demonstrate that this is a folded domain that has been "trapped" by the presently described methods.

EXAMPLE 6**COMPARISON OF THE NMR STRUCTURE OF CSP A
WITH OTHER PROTEINS**

Recombinant CspA is expressed and purified using the protocol essentially as described by Chatterjee *et al.*, *J. Biochem.* 114:663-669 (1993), and Feng *et al.*, *Biochemistry* 37:10881-10896 (1998), both of which are incorporated by reference in their entirety. The purified CspA protein is prepared for NMR analysis by dialysis against a buffer containing 50 mM potassium phosphate and 1 mM NaN_3 , pH 6.0 and the sample is analyzed using a Varian Unity 500 spectrometer equipped with three channels and a fourth frequency synthesizer for carbonyl decoupling as described by Feng *et al.*, *Biochemistry* 37:10881-10896 (1998). Figure 7 provides the 2D ^{15}N - ^1H HSQC spectrum of CSP A at pH 6.0 and 30°C.

The collected spin resonances are analyzed using AUTOASSIGN. The input for AUTOASSIGN includes peaks from 2D ^{15}N - ^1H HSQC and 3D HNCB spectra along with peak lists from three intrareidue (CANH, CBCANH and HCANH) and three interresidue (CA(CO)NH, CBCA(CO)NH and HCA(CO)NH) experiments, which correlate with the C^α , C^β and H^α resonances of residues *i* and *i*-1 respectively. The results of the AUTOASSIGN analysis of the peak picked 2D and 3D NMR spectra are summarized in Table 1.

Side chain resonance assignments are obtained using PFG HCCNH-TOCSY and PFG HCC(CO)NH-TOCSY and homonuclear TOCSY experiments recorded with multiple mixing times of 22, 36, 45, 54, 71 and 90 ms according to the method of Celda

and Montelione, *J. Magn. Reson. Biol.* 189-193 (1993), incorporated by reference in its entirety. Interatomic distance constraints are derived from three NOESY data sets 2D NOESY and 3D ^{15}N -edited NOESY-HSQC spectra recorded with a mixing time of t_m of 60 ms of a CspA sample dissolved in 90% H_2O /10% $^2\text{H}_2\text{O}$ and a 2D NOESY spectrum is recorded with a mixing time t_m of 50 ms of a sample dissolved in 100% $^2\text{H}_2\text{O}$. The intensity of the NOESY-HSQC spectrum is corrected for ^{15}N relaxation effects, and the cross-peak intensities are converted into interproton distance constraints.

Table 1					
Summary of AUTOASSIGN Analysis for CspA Triple-Resonance NMR Data					
Residues	69		Number of assignments (expected)	AUTOASSIGN analysis	Manual analysis
			Backbone		
GSs expected	66		H^{N}	65	66
GSs observed	67		H^{a}	77	79
Degenerate GS roots	8		^{15}N	65	66
Assigned GSs	65		$^{13}\text{C}^{\text{a}}$	67	69
Extra GSs	2		$^{13}\text{C}^{\text{b}}$	64	66
Assigned residues	68		$^{13}\text{C}^{\text{b}}$	49	59
Percent assigned residues	99%		Side chain		
Execution times (sec.)	10		^{15}N	6	6
			H^{N}	11	11

Stereospecific assignments of methylene H^{b} s are made by analysis of local NOE and vicinal coupling constant data using the HYPER program. HYPER is a conformational grid search program used for determining stereospecific $\text{C}^{\text{b}}\text{H}_2$ methylene proton assignments and for defining the ranges of dihedral angles ϕ , ψ , χ^1 that are consistent with the local experimental NMR data for each amino acid in a polypeptide (Tejero *et al.*, *J. Biomol. NMR* (in press), incorporated by reference in its entirety). The secondary structural elements of CspA are summarized in Figure 8. From this information, five β -strands corresponding to polypeptide segments of residue 5-13, 18-22, 30-33, 50-56 and 63-70 are identified.

The average number of distance constraints per residue is 10.4. Dihedral angle constraints are obtained from the HYPER program. Structure generation calculations are carried out with DIANA, version 2.8 (TRIPOS, Inc.) using R8000 processor in a Silicon Graphics Onyx workstation (Braun and Go, *J. Mol. Biol.* 186:611-626 (1985), and Guntert *et al.*, *J. Mol. Biol.* 169:949-961 (1983), both of which are incorporated by reference in their entirety).

From this NMR data set, the solution structure of CspA is reasonably well defined. Using the refined CspA coordinates defined by the present invention, structural database searches of the Protein Data Base (PDB) are performed with the DALI program. This search is able to identify a list of proteins or domains of structural homologues. Identified structural homologues of CspA exhibiting similar biochemical function include the RNA binding domain of *E. coli* polyribonucleotide nucleotidyltransferase, the human mitochondrial ssDNA-binding protein, *E. coli* translation initiation factor 1, the ssDNA-binding protein from gene V of filamentous bacteriophages M13 and f1, the ssDNA-binding protein from *Pseudomonas* phage Pf3, elongation factor G from *Thermus thermophilus*, a domain of *E. coli* lysyl tRNA synthetase, a domain of yeast tRNA synthetase, human replication protein A, staphylococcus nuclease, and a domain of *E. coli* topoisomerase I. Although the function of CspA was already known, the present Example has illustrated the use of the present invention.

As the present invention describes, a person of skill in the art is able to take a polypeptide of unknown function, express and purify a stable peptide domain encoded by the polypeptide, determine the NMR 3D structure of that expressed domain and predict the function of that domain by comparing the structure of that domain against known structures having known functions. This represents a fundamental paradigm shift in the study of proteins.

EXAMPLE 7

AUTOMATED ANALYSIS OF PROTEIN STRUCTURES FROM NMR DATA

Figure 9 outlines the constraint reasoning system of the present invention which automatically generates protein structures from NMR data. Briefly, the constraint reasoning system is based on automated analysis of secondary structure, prediction of hydrophobic core contacts, and iterative analysis of contact frequencies. The constraint reasoning generates reliable initial chain folds even when the chemical shift information alone provides few unambiguous NOESY cross peak assignments.

In the first step, a Simple Match is performed to determine all possible assignments (A-type matches) for each spectra. In the second step, the expected peaks which are consistent with secondary structure, or which are intra/seq are identified. These peaks are placed in an experimental (E) and an unknown (U) set. The expected peaks are further used to create a dynamically locally referenced values (DLRV) for H and HX (local referencing). The DRLV for each atom in each dimension includes the

original chemical shift value plus any additional chemical shift values derived from the E set. If only one expected match is found for a given peak, that peak is put into U and E set. If more than one expected match is found for a given peak (B-type expected matches), those expected matches are also put into U and E set.

In the third step, the local match tolerance for HX dimension is defined. The local match tolerance for HX dimension is based on assigned HX resonance from E set. HX resonance is performed as described by Koide *et al.*, *J. Biomol. NMR* 6:306-312 (1995); Bai *et al.*, *Proteins* 20:4-14 (1994); and Englander and Mander, *Annu. Rev. Biophys. Biomol. Struct.* 21:243-265 (1992), all of which are incorporated by reference in their entirety.

In the fourth step, U peaks are supplemented based on chemical shift (unambiguous) data filtered through a noise filter. The noise filter reduces the background noise by eliminating peaks having an intensity of <0.05% of the highest intensity of the real intra peaks. Thus, a tighter match tolerance to chemical shift list is created by the noise filter makes than the list created by the Simple Match of step 1.

B-type matches, a subset of A-type matches for each spectra, are defined in step 5. The B-type matches for a given peak are defined by ordering the A-type matches based on the size of the match value. The match value is computed as follows:

$$MV = \min(\Delta HX + \Delta X/10 + \Delta H)$$

where $\Delta H = H_{obs} - H_{DCSL}$; $\Delta HX = HX_{obs} - HX_{DCSL}$; $\Delta X = X_{obs} - X_{DCSL}$; and H_{DCSL} , HX_{DCSL} and X_{DCSL} are sets of dynamically locally referenced values (DLRV) for the H, HX, and X dimensions, respectively. All possible matches with $\gamma \leq 0.01$ are chosen, where $\gamma = \frac{1}{MV} - (\Delta HX + \Delta X/10 + \Delta H)$.

In step 6, the Contact Frequency (CF) of E is used to assign B-type matches to U set. A contact bin is created from all E's. If a peak in B is in the contact bin, it is assigned to U. Otherwise, it is assigned to T-type matches. In step 7, SYM, a constraint satisfaction program, is used to assign B-type matches to U set. If a peak in B has symmetry to another peak in B, both are assigned to U set as T-type assignments. SYM modeling is performed utilizing the method described by Gdaniec *et al.*, *Biochemistry* 37:1505-1512 (1998); Easterwood and Harvey, *RNA* 3:577-585 (1997); Laing and Hall, *Biochemistry* 35:13586-13596 (1996); Ericson *et al.*, *J. Mol. Biol.* 250:407-419 (1995); and Foucrault and Major, *ISMB* 3:121-126 (1995), all of which are incorporated by reference in their entirety. In step 8, HP-CORE, which predicts buried residues, is used to assign B-type matches to U set. A HP-CORE contact bin is created from all B's. If

the contact frequency (CF) of the HP-CORE contact bin is $> N$, all peaks in this bin are assigned to U as T-type assignments. N is a heuristic value that is scale with the number of NOESY spectra available.

The 3D structure of the protein is computed in step 9. First, the structure calculation program is calibrated, where the distance of D-type peaks are derived from their intensity and the distance of T-type peaks are $= 5.0\text{\AA}$. The structure calculation program is then run. The 10 best results, from a family of 50 3D structures are selected. For each of the 10 best results, the $S(\phi)$, $S(\varphi)$, $\sigma(i,j)$ matrix, bb root mean square deviation (RMSD) are calculated where records with a $S(\phi) < 0.7$ and $S(\varphi) < 0.7$ are excluded. If the rmsd is too large, further analysis is stopped. If the rmsd is $< 1\text{\AA}$, the analysis continues with step 12. If it has progress, analysis continues with step 10. If there isn't any more progress, analysis proceeds with the next cycle (decrease O). Disordered regions – order (i,j) are identified from O. If $(\langle S - O \rangle \geq 0$ and $(\sigma(i,j) - 2/O) \leq 0)$ and $\text{order}(i,j) = 1$, then the region is ordered. If $\text{order}(i,j) = 0$, then the region is disordered.

In the validation step, step 10, peaks that consistently violated NOE assignments are removed from U list. If the peak is greater than the Violation Parameter (V), it is assumed that the assignment is wrong. If $\text{order}(i,j) = 1$, then $V = 1$ and if $\text{order}(i,j) = 0$, then $V = 2$. If the $v_{\min}(i,j)$ is greater than V and it is a T-type assignment, it is deleted from the assignment list. If it is a D-type assignment, it is downgraded to a T-type assignment and assigned an alternate assignment of $\langle d \rangle < 5\text{\AA}$. If a peak has more than one T-type assignment and only one of the peaks has violated V, it is reassigned as a D-type assignment.

In step 11, expected peaks that are consistent with 3D structure are identified and placed in U set. It is assumed that if the peak is in an ordered region and it is greater than the Distance Cutoff (D), it is an incorrect assignment. If $(\text{order}(i,j) = 1)$, then $D = 5 + \text{rmsd} * 2$ and $D_{\min} = 5.5\text{\AA}$. N , the number of possible assignments left, is put into U set. If $\text{rmsd} > 2$, then $N \leq 2$. If $\text{rmsd} > 1$, then $N \leq 3$. For any other rmsd value, $N \leq 4$. Any assignment with a $d_{\min}(i,j) > D$ in ordered region is removed from A list. If N possible assignments are left, they are put into U set as T-type assignments.

In set 12, all possible NOE's that are expected from the structure are back calculated. Any predicted assignments not in U or A list and any peak still in A list are outputted. For each cycle, a Contact Map (assignment, structure), Connectivity Map, Structures, Assignments (ordered by intra, seq, mid, long range), $S(\phi)$, $S(\varphi)$, $\sigma(i,j)$ matrix, and bb rmsd are outputted.

EXAMPLE 8

AUTOMATED GENERATION OF 3D STRUCTURES

The constraint reasoning system, outlined in Figure 9 and described in Example 7, is used to automatically generate the 3D structures of the Zdom and Cspa proteins (Figures 10A and B, and Figure 17, respectively). The constraint reasoning system generated 3D structures are compared to the manually generated 3D structures. The results of the automated assignment analysis for Zdom and Cspa are presented in Figures 11-13 and 18-20, respectively. The results of the manual assignment analysis for Zdom and Cspa are presented in Figures 14-16 and 21-23, respectively. Backbone – backbone assignments are designated by x. Backbone – side chain assignments are designated by o. Side chain – side chain assignments are designated by □. Intra-residue assignments are designated by filled symbols.

In a further embodiment, a constraint reasoning system for automatically generating protein structures from NMR data is employed. A variety of constraints have been used to resolve the ambiguity problem in analysis of 2D and 3D NOESY spectra, obtain an initial chain fold, and then use constraints implied by this initial structure to iteratively refine the protein structure. The constraint reasoning system is based on automated analysis of secondary structure, prediction of hydrophobic core contacts, and iterative analysis of contact frequencies. The constraint reasoning system can generate reliable initial chain folds even when the chemical shift information alone provides few unambiguous NOESY cross peak assignments. Experimental NMR data for two different proteins have been analyzed to automatically generate 3D structures. The structures generated by this constraint reasoning system in hours are in good agreement with those derived from manual analysis processes which require weeks or months.

The NOESY-Assign constraint reasoning system for this purpose comprises the following 12 steps:

Step 1: Simple Match – get all possible assignments (A-type matches) for each spectra.

Step 2: Identify expected peaks which are intra/seq, or consistent with secondary structure. Put in U and E set. Create dynamically referenced values (DLRV) for H and HX (local referencing). The DLRV for each atom in each dimension includes the original chemical shift value plus any additional chemical shift values derived from E set.

Given a peak, if only one expected match is found, put in U and E set.
If found more than one expected match is found, select B-type
expected matches, put in U and E set. See Step 5 for explanation of
B-type match.

* Not for 2D spectra

- All assignments are D-type assignments
- Remove all that are inconsistent with secondary structure

*** Possible features ***

1. Check if the data set are consistent with each other
 - List the residue that no intra HN – Ha in N15-NOESY
 - List the residue that no intra Ha – Hb in C13-NOESY
 If have, let the user do local re-referencing or global re-referencing
2. Do referencing refinement

Step 3: Define local match tolerance for HX dimension based on assigned HX
resonance from E set.

* Not for 2D spectra

Define local match tolerance for HX dimension:

For each possible HX dimension assignment, find all assignments
in E set, calculate the 60% confidence region. If the peak's
chemical shift in the HX dimension is outside of the 60%
confident region (using common sample statistics methods),
remove it from the list of possible assignments (flag).

Step 4: Supplement U based on chemical shift (unambiguous) with noise
filter.

Noise filter: Basic idea is that real peaks have:

- intensity > 0.05% of the highest intensity of real peaks
- tighter match tolerance to chemical shift list than used in Step 1
(Simple Match)

Highest intensity of real peaks – what is real peaks? Use the highest
intensity of intra peaks.

T-type assignments

Step 5: Define B-type matches, subset of A-type matches for each spectra.
The B-type matches are defined as follows (by default):

For a given peak, order the A-type matches based on the size of the match value (MV), which is computed as follows:

$$MV = \min(\Delta HX + \Delta X/10 + \Delta H)$$

$$\text{where: } \Delta H = H_{\text{obs}} - H_{\text{DCSL}}$$

$$\Delta HX = HX_{\text{obs}} - HX_{\text{DCSL}}$$

$$\Delta X = X_{\text{obs}} - X_{\text{DCSL}}$$

and H_{DCSL} , HX_{DCSL} , and X_{DCSL} are the sets of dynamically locally referenced values (DLRV) for the H, HX and X dimensions, respectively. Choose all possible matches with: $\gamma \leq 0.01$, where $\gamma = |MV - (\Delta HX + \Delta X/10 + \Delta H)|$.

Step 6: Use Contact Frequency (CF) of E to assign B-type matches to U set

* Not for 2D spectra

- Create contact bin from all E's
- If element in B is in contact bin, Assign to U
- T-type assignment

Step 7: Use SYM (Symmetry Property) to assign B-type matches to U

* Not for 2D spectra

If peak in B has another symmetry peak in B, Assign both to U, as T-type assignments

Step 8: Use HP-CORE to assign B-type matches to U

* Not for 2D spectra

HP-CORE: Predicted Buried Residue

- Create HP-CORE contact bin from all B's
 - HP-CORE to HP-CORE
 - not in the same secondary segment
- If CF of the HP-CORE contact bin > N, assign all peaks in this bin to U, as T-type assignments. N is heuristic value that should scale with the number of NOESY spectra available, a typical value of N is 2.
- If element in B is in contact bin, Assign to U
- T-type assignment

Step 9: Compute 3D structure

O: Order Parameter

0.8 (cycle 1), 0.7 (cycle 2), 0.6, (cycle 3), 0.5 (cycle 4)

1. Calibration

- D-type: distance is derived from its intensity
- T-type: distance = 5.0 Å

2. Run Structure Calculation Software

3. Select 10 best, from family of 50 3D structures:

- Compute: $S(\phi)$, $S(\varphi)$, $\sigma(i,j)$ matrix, bb rmsd (exclude record with $S(\phi) < 0.7$ and $S(\varphi) < 0.7$)
- if bb rmsd is too large, STOP
- if bb rmsd is < 1 Å, go to step 12

4. If has progress, go to step 10

5. If no more progress, decrease O (next cycle). Identify disordered regions – order(i,j), from O

If $(\langle S - O \rangle \geq 0 \text{ \& } \sigma(i,j) - 2/O \leq 0$

Order (i,j) = 1 (ordered)

Else Order (i,j) = 0 (disordered)

Example:

Cycle1: $O = 0.8$, $2/O = 2.5$

Cycle2: $O = 0.7$, $2/O = 2.85$

Cycle3: $O = 0.6$, $2/O = 3.33$

Cycle4: $O = 0.5$, $2/O = 4$

Step 10: Validation – remove from U list that consistently violated NOE assignments

V: Violation Parameter

Assumption: if $> V$, for sure, it is wrong assignments

If order(i,j) = 1, $V = 1$,

If order(i,j) = 0, $V = 2$

If $vmin(i,j) > V$

- T-type: Delete it from the possible assignment list
- D-type: Downgrade to T-type assignment, and assign alternate assignments of $\langle d \rangle < 5$ Å also as T-type assignments
- If a peak has more than one T-type assignments,
If only one that is not violated, make it as D-type assignments

Step 11: Identify expected peaks that are consistent with 3D structure, put in U set.

D: Distance Cutoff

Assumption: If in ordered region, and $> D$, for sure, that is impossible to be a right assignment

If (order(i,j) = 1)

$D = 5 + \text{rmsd} * 2$ and $D_{\min} = 5.5 \text{ \AA}$

N: Number of possible assignments left and put in U.

If $\text{rmsd} > 2$, $N \leq 2$, If $\text{rmsd} > 1$, $N \leq 3$,

Rest, $N \leq 4$,

Pruning A list:

- Remove possible assignment with $\text{dmn}(i,j) > D$ in ordered region
- If N possible assignment left, put in U as T-type assignments

Step 12: Back calculate all possible NOE' that are expected from the structure. Output any predicted assignments not in U or A list and peaks still in A list.

Output:

For each cycle: Contact Map (assignment, structure(), Connectivity Map, Structures, Assignments (ordered by intra, seq, mid, long range), $S(\phi)$, $S(\varphi)$, $\sigma(i,j)$ matrix, bb rmsd

Overview:

- Number of Assignments for each assignment step
- | Table | #Total NOE | #U(D+T) | #A | #Noise |
|-------|------------|---------|----|--------|
| | | | | |
- Noise Peak List
- A-type matches List

It will be apparent to those skilled in the art that various modifications may be made in the present invention without departing from the spirit and scope of the present invention. It will be additionally apparent to those skilled in the art that the basic construction of the present invention is intended to cover any variations, uses or adaptations of the invention following, in general, the principle of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains. Therefore, it will be appreciated that the scope of this invention is to be defined by the claims appended hereto, rather than the specific embodiments which have been presented as examples.

Claims

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WHAT IS CLAIMED IS:

1. A high-throughput method for determining a biochemical function of a protein or polypeptide domain of unknown function comprising:
 - (A) identifying a putative polypeptide domain that properly folds into a stable polypeptide domain, said stable polypeptide having a defined three dimensional structure;
 - (B) determining three dimensional structure of the stable polypeptide domain from an automated analysis of NMR spectrometer spectra of said polypeptide domain, wherein said automated analysis is conducted by a NOESY_Assign process;
 - (C) comparing the determined three dimensional structure of the stable polypeptide domain to known three-dimensional structures in a protein data bank, wherein said comparison identifies known structures within said protein data bank that are homologous to the determined three dimensional structure; and
 - (D) correlating a biochemical function corresponding to the identified homologous structure to a biochemical function for the stable polypeptide domain.
2. The method according to claim 1, further comprising the prestep of parsing a target polynucleotide into at least one putative polypeptide domain.
3. The method according to claim 2, wherein said parsing is performed by a first computer algorithm, wherein said first computer algorithm is selected from the group consisting of a computer algorithm capable of determining exon phase boundaries of a polynucleotide, and a computer algorithm capable of determining interdomain boundaries encoded in a polynucleotide.
4. The method of claim 3, further comprising a computer algorithm that compares the putative polypeptide domain sequence with known domain sequences stored within a database.
5. The method of claim 1, wherein said NMR spectra are analyzed by a second computer algorithm that automatically assigns resonance assignments to the polypeptide sequence.

- 5
6. The method of claim 1, wherein said identification of said stable polypeptide domain comprises measuring a time course of amide hydrogen-deuterium exchange.
- 10
7. The method of claim 1, wherein prior to step (B), said stable polypeptide domain is optimally solubilized, said optimum solubilization comprising:
- 5
- i) preparing an array of microdialysis buttons, wherein each of said microdialysis buttons contains at least 1 μ l of an approximately 1mM solution of said stable polypeptide domain;
 - 15 ii) dialyzing each member of said array of microdialysis buttons against a different dialysis buffer;
 - 10 iii) analyzing each of said dialyzed microdialysis buttons to determine whether said stable polypeptide domain has remained soluble; and
 - 20 iv) selecting the polypeptide domain having optimum solubility characteristics for NMR spectroscopy.
- 25
8. The method of claim 1, wherein said comparison of said determined three dimensional structure to said known three-dimensional structures in the protein data bank is performed by a third computer algorithm that is capable of determining 3D structure homology between said determined three dimensional structure and a member of said PDB.
- 30
9. The method according to claim 11, wherein said third computer algorithm is selected from the group consisting of DALI, CATH and VAST.
- 35
10. The method of claim 1, wherein said protein data bank is Protein Data Base ("PDB").
- 40
11. The method of claim 4, wherein said database contains domain sequence information of known and determined domain sequences.
- 45
12. An integrated system for rapid determination of a biochemical function of a protein or protein domain of unknown function:
- (A) a first computer algorithm capable of parsing said target polynucleotide into at least one putative domain encoding region;
 - 30 (B) a designated lab for expressing said putative domain;
 - (C) an NMR spectrometer for determining individual spin resonances of amino acids of said putative domain;
- 50
- 55

- 5 (D) a data collection device capable of collecting NMR spectral data,
wherein said data collection device is operatively coupled to said NMR
spectrometer;
- 10 (E) at least one computer;
- 5 (F) a second computer algorithm capable of assigning individual spin
resonances to individual amino acids of a polypeptide;
- 15 (G) a third computer algorithm capable of determining tertiary structure of a
polypeptide, wherein said polypeptide has had resonances assigned to
individual amino acids of said polypeptide;
- 10 (H) a database, wherein stored within said database is information about the
structure and function of known proteins and determined proteins; and
- 20 (I) a fourth computer algorithm capable of determining 3D structure
homology between the determined three-dimensional structure of a
polypeptide of unknown function to three-dimensional structure of a
15 protein of known function, wherein said protein of known structure is
stored within said protein database, wherein said fourth computer
25 algorithm determines said structure by an automated NOESY_Assign
process.
13. A high-throughput method for determining a biochemical function of a
20 polypeptide of unknown function encoded by a target polynucleotide comprising
the steps:
- 30 (A) identifying at least one putative polypeptide domain encoding region of
the target polynucleotide ("parsing");
- (B) expressing said putative polypeptide domain;
- 35 25 (C) determining whether said expressed putative polypeptide domain forms a
stable polypeptide domain having a defined three dimensional structure
("trapping");
- (D) determining the three dimensional structure of the stable polypeptide
40 domain by an automated NOESY_Assign process;
- 30 (E) comparing the determined three dimensional structure of the stable
polypeptide domain to known three dimensional structures in a Protein
Data Bank to determine whether any such known structures are
45 homologous to the determined structure; and
- 35 (F) correlating a biochemical function corresponding to the homologous
structure to a biochemical function for the stable polypeptide domain.

FIGURE 1

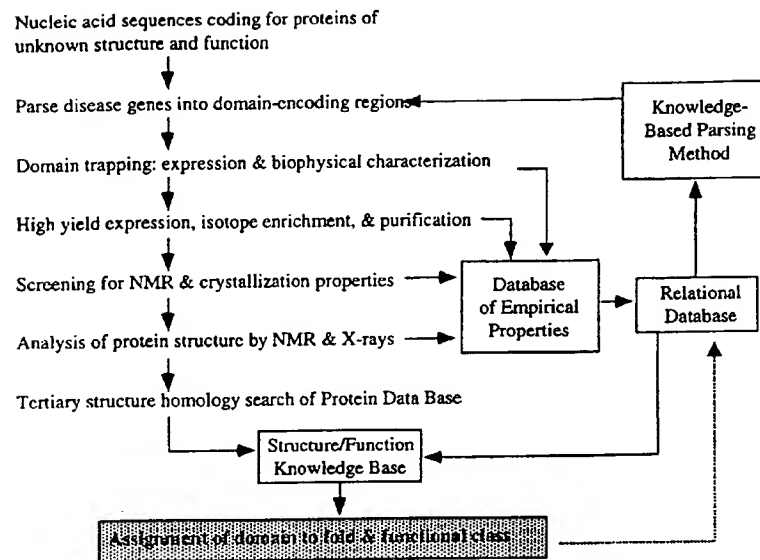


FIGURE 2A

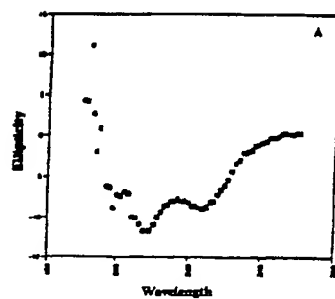


FIGURE 2B

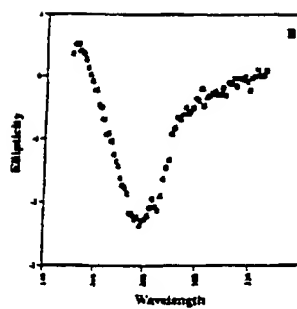


FIGURE 3

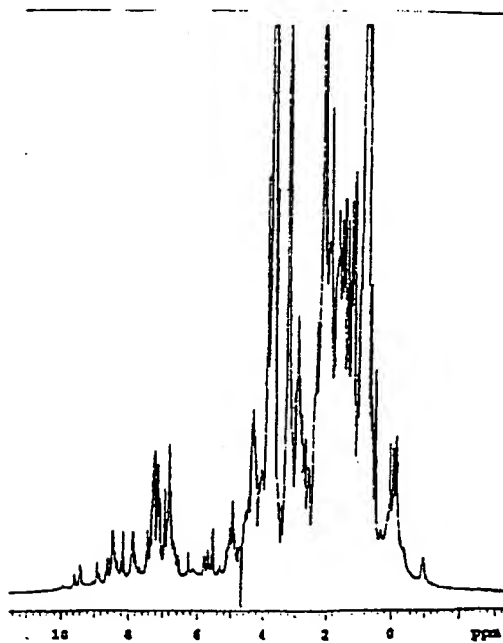


FIGURE 4

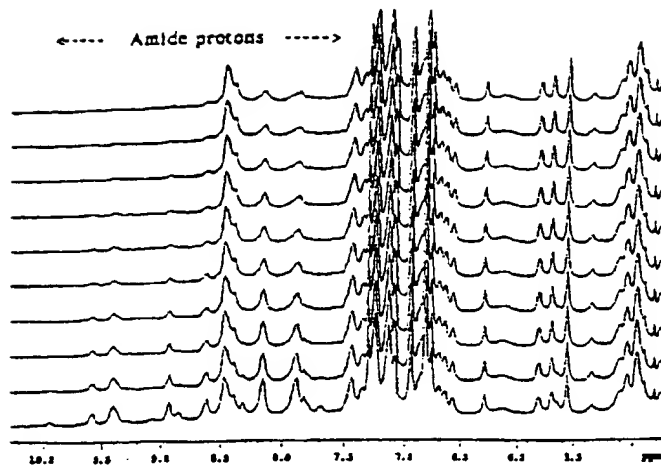
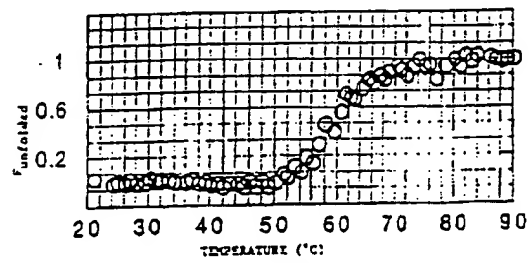


FIGURE 5



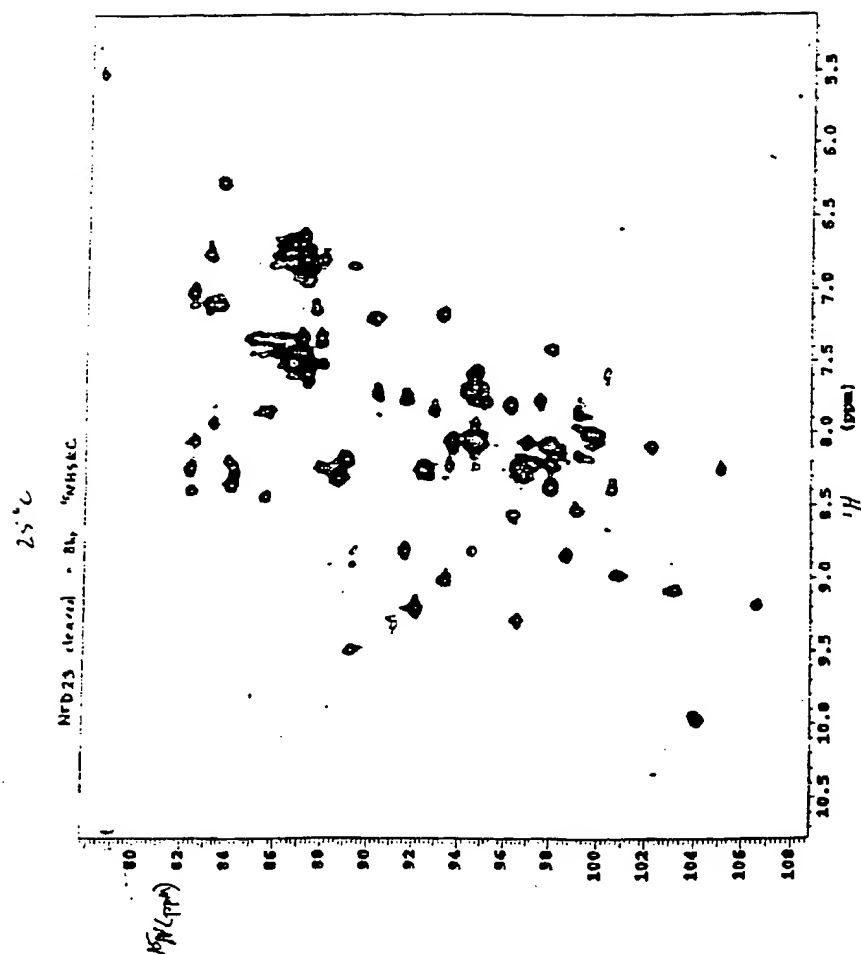


FIGURE 6

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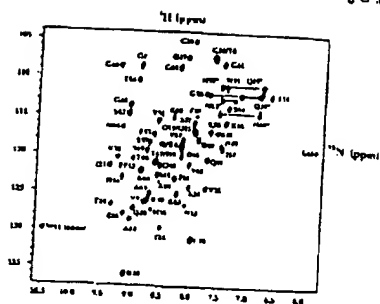
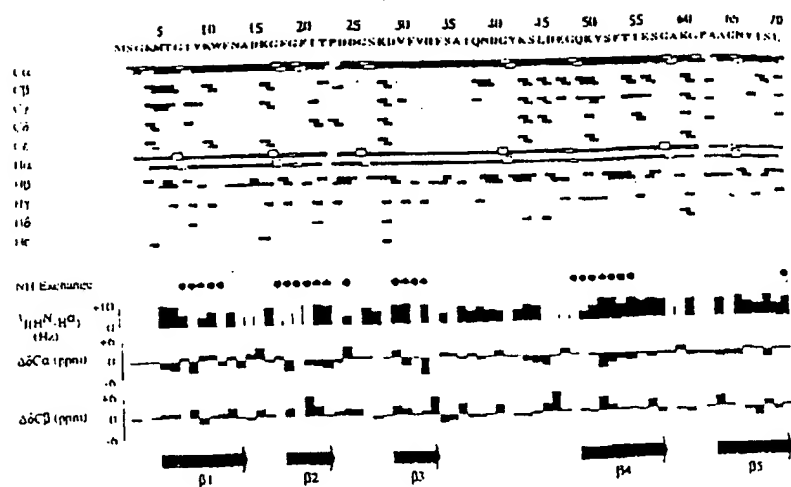


Figure 7



Figures 8A+B

Figure 9:
NOESY_ASSIGN Process

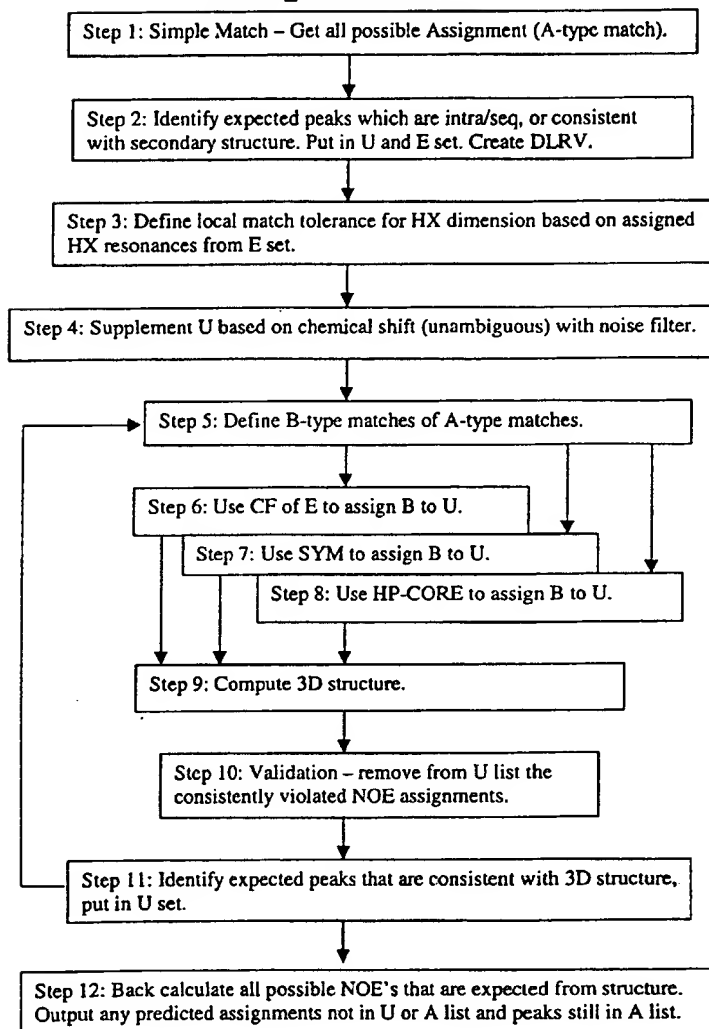
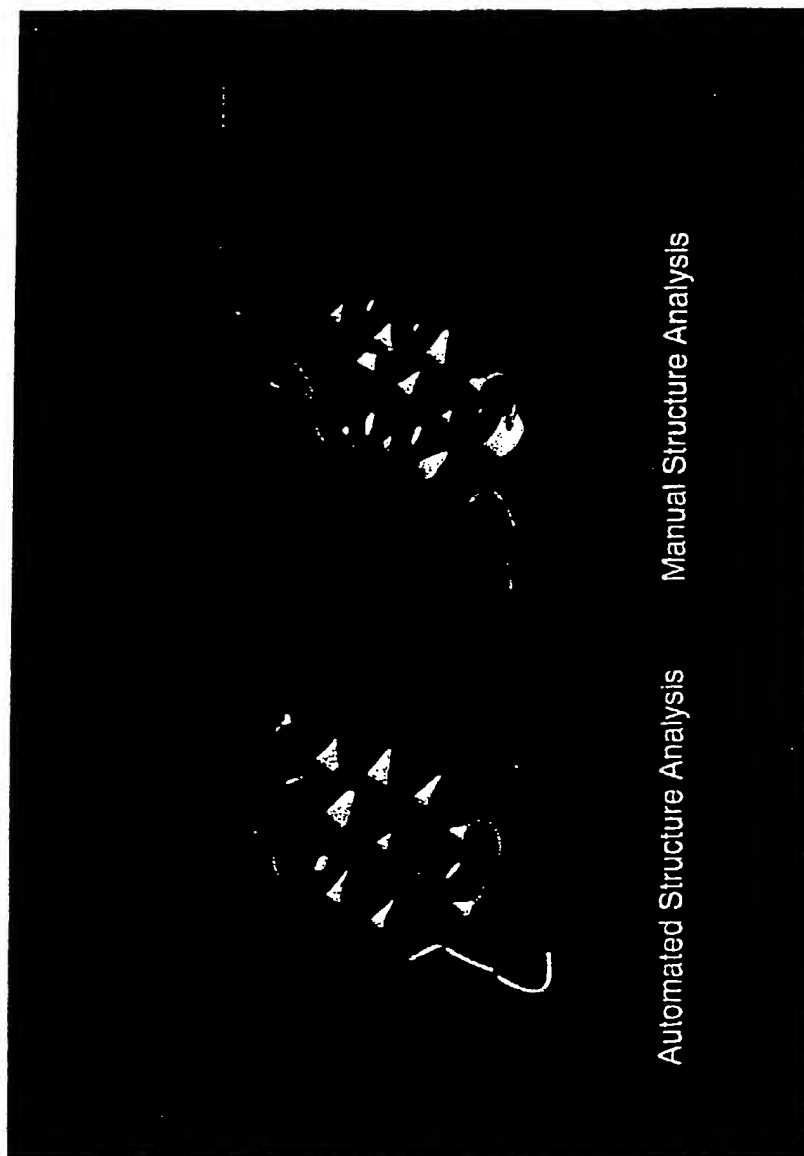
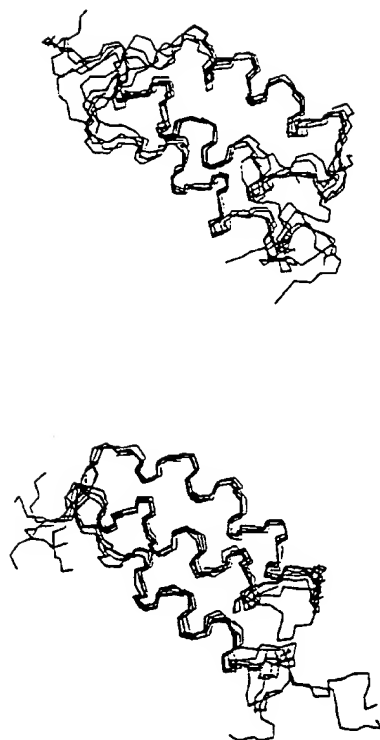


Fig 10 A





Manual Structure Analysis

Automated Structure Analysis

Fig 10B

Zdom: Automated Assignment Analysis

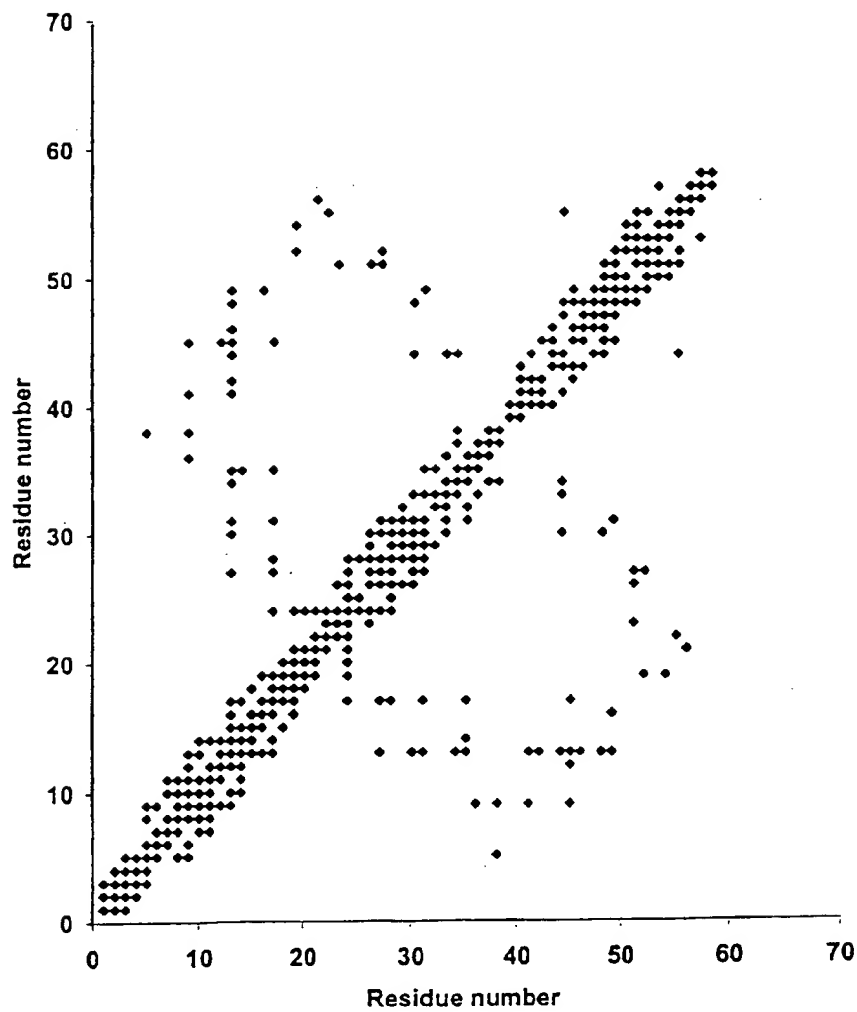


Fig. 11

Zdom: Automated Structure Analysis

Fig. 12

- × backbone - backbone (filled diamond if intraresidue)
- backbone - side chain (filled if intraresidue)
- side chain - side chain (filled if intraresidue)

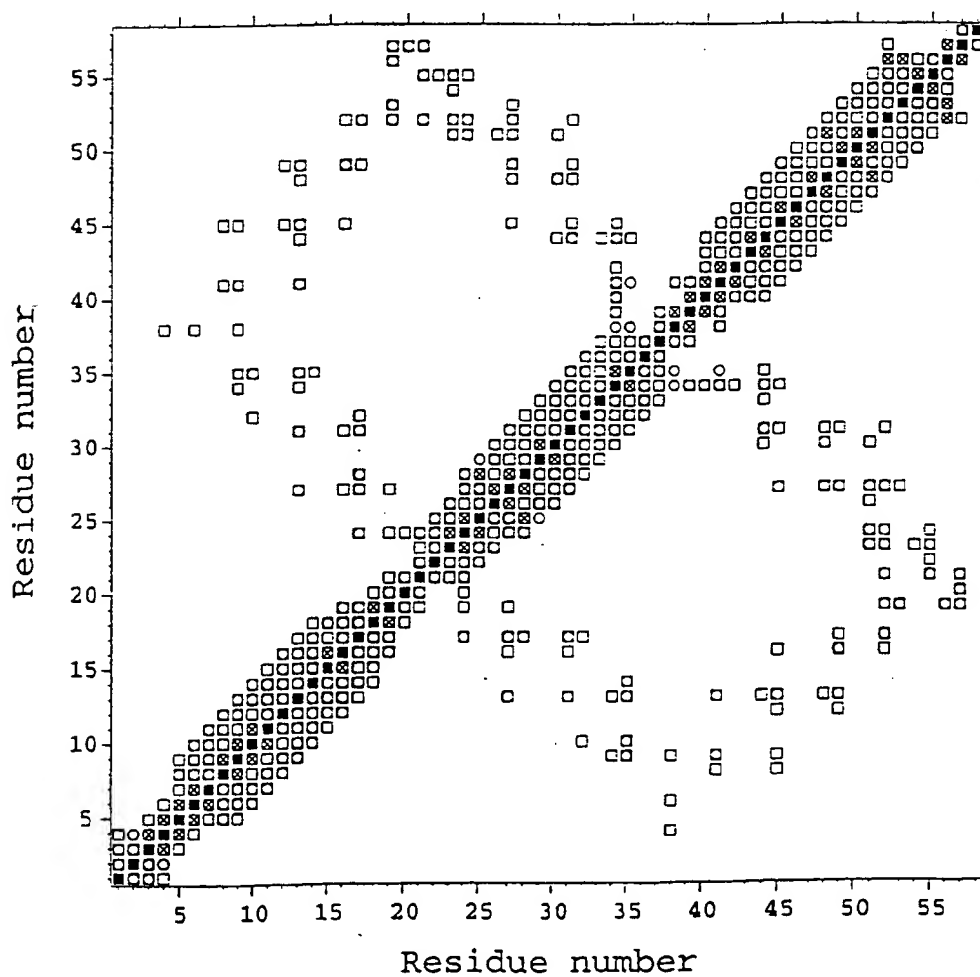
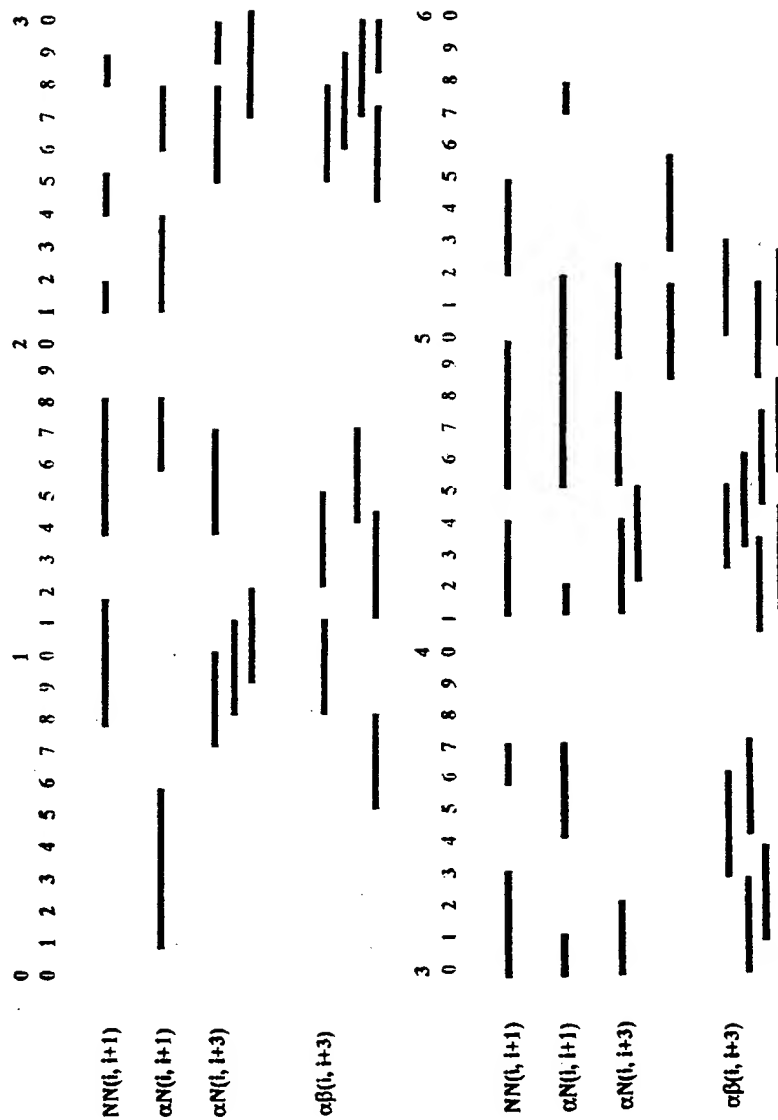


Fig. 13

Zdom: Automated Analysis — Connectivity Map



Zdom: Manual Assignment Analysis

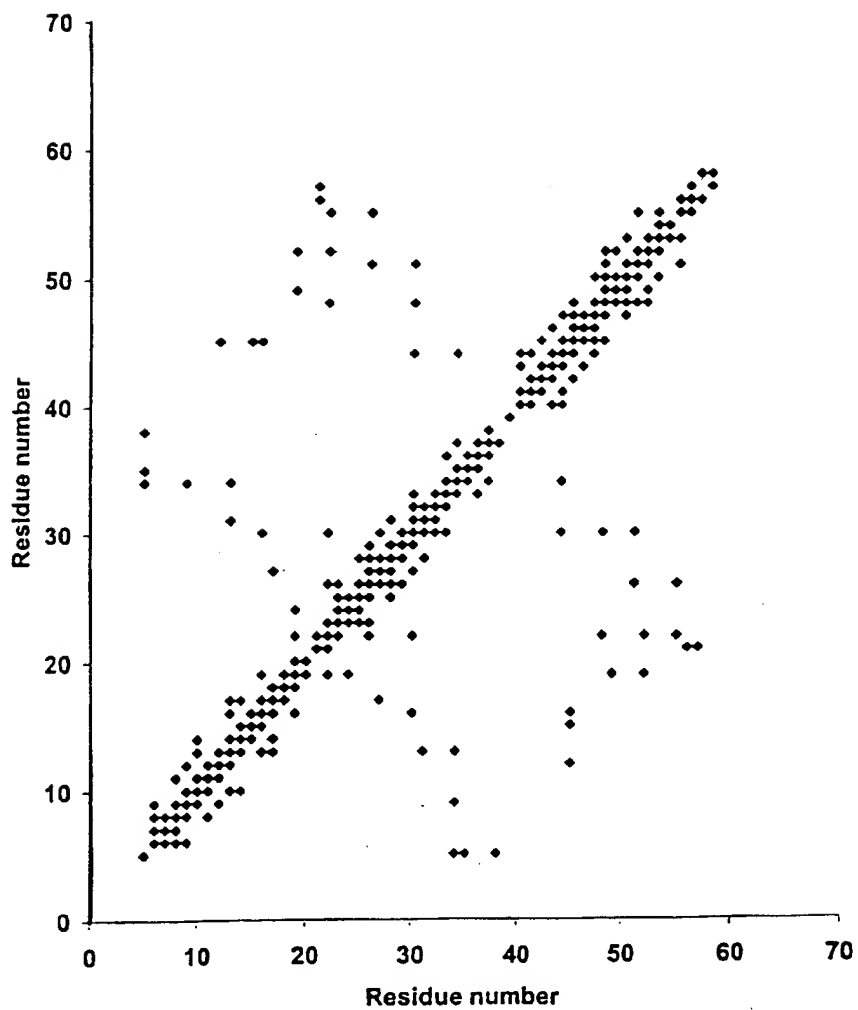


Fig 14

Zdom: Manual Structure Analysis

- × backbone - backbone (filled diamond if intraresidue)
- backbone - side chain (filled if intraresidue)
- side chain - side chain (filled if intraresidue)

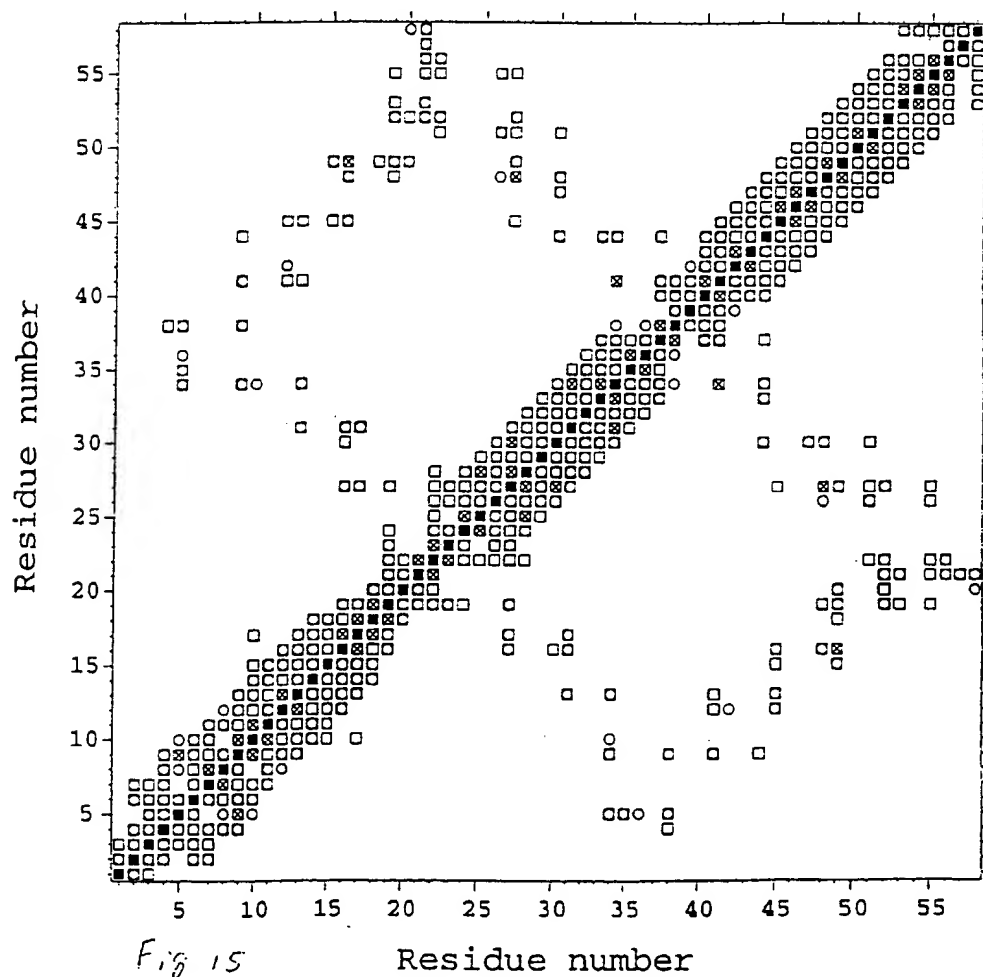
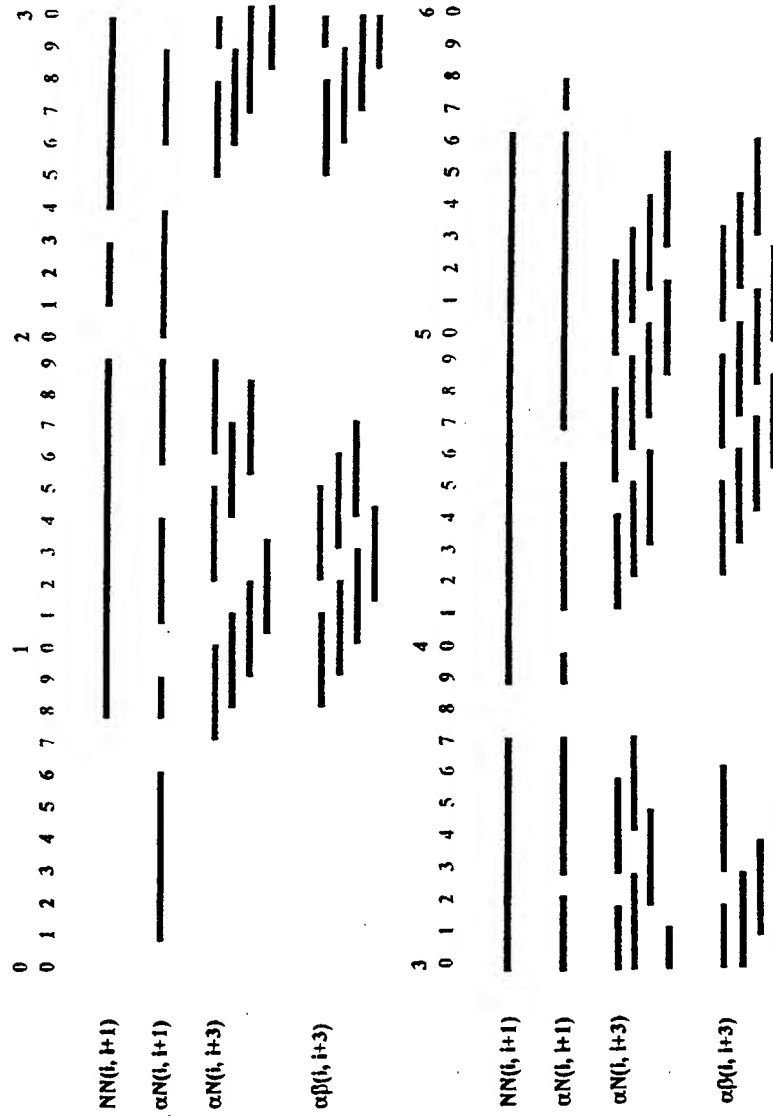
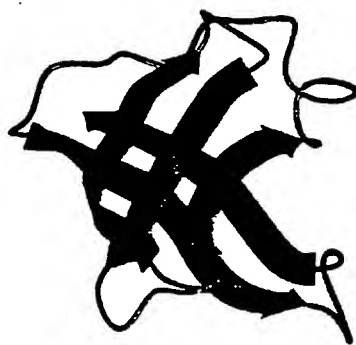


Fig: 16
Zdom: Manual Analysis—Connectivity Map



CspA



Automated Structure Analysis



Manual Structure Analysis

Fig 17

Cspa: Automated Assignment Analysis

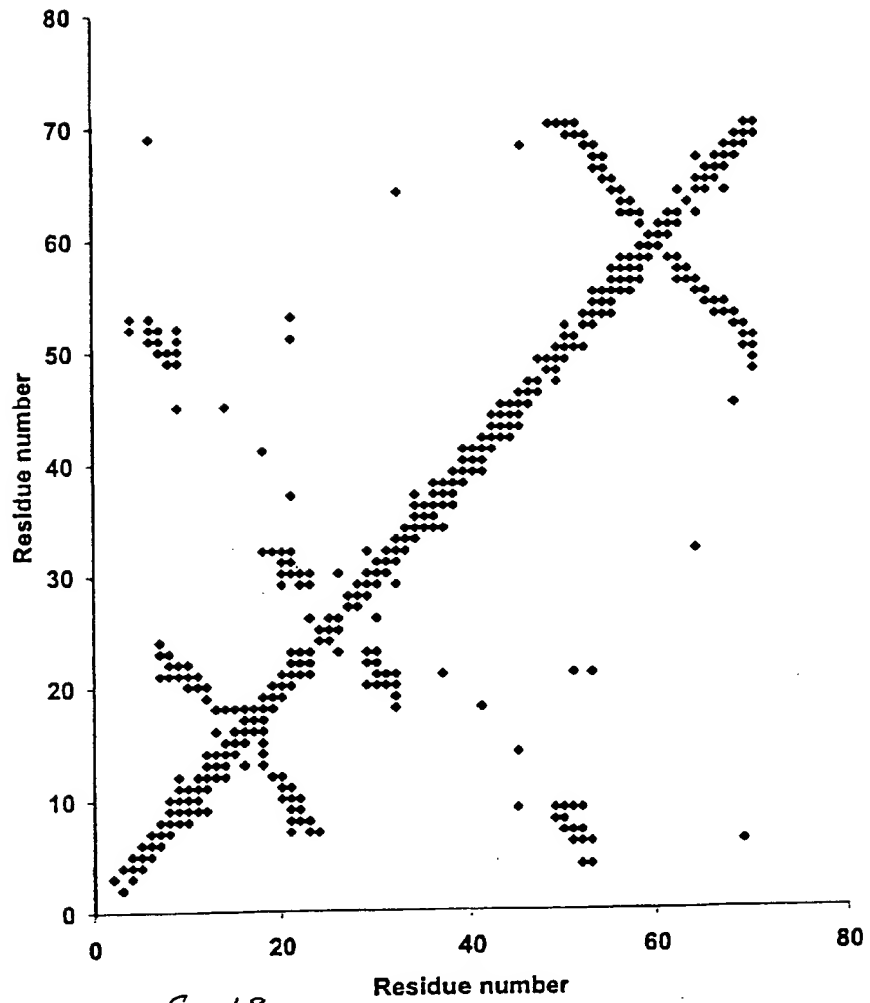
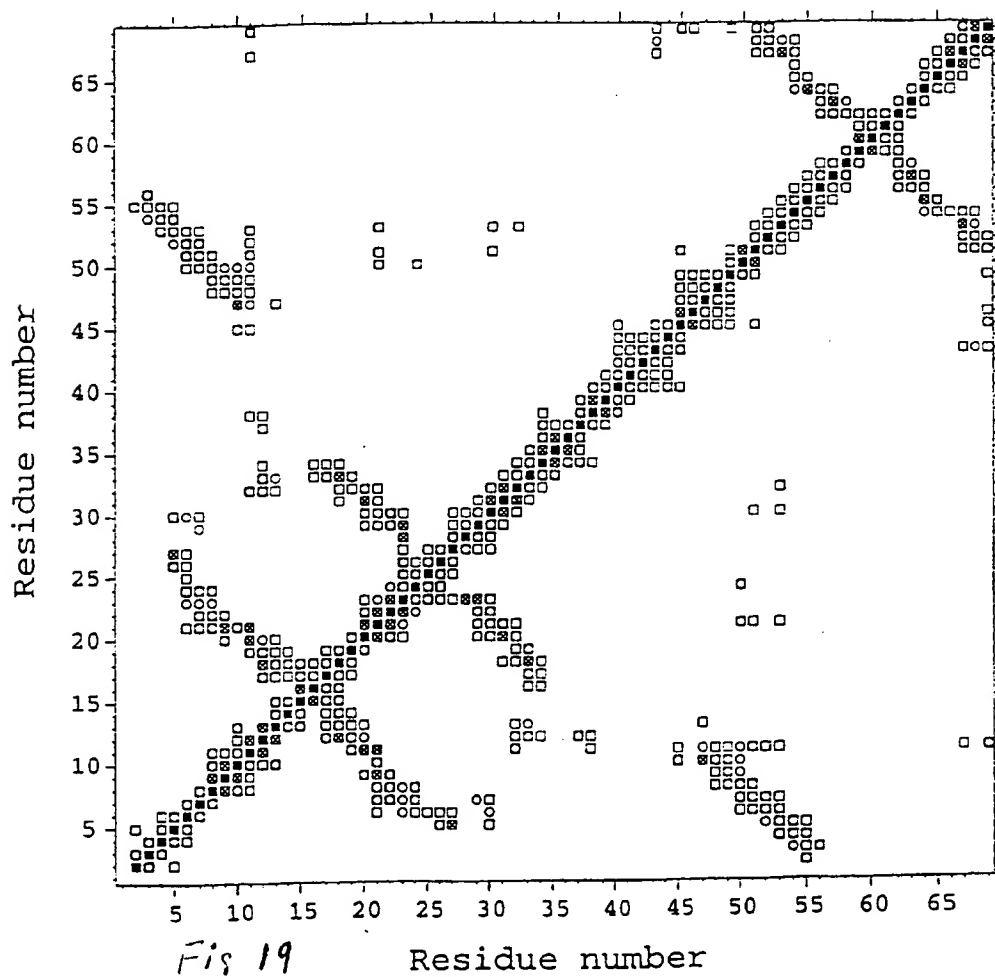


Fig 18

Cspa: Automated Structure Analysis

- × backbone - backbone (filled diamond if intraresidue)
- backbone - side chain (filled if intraresidue)
- side chain - side chain (filled if intraresidue)



Cspa: Automated Analysis — Connectivity Map

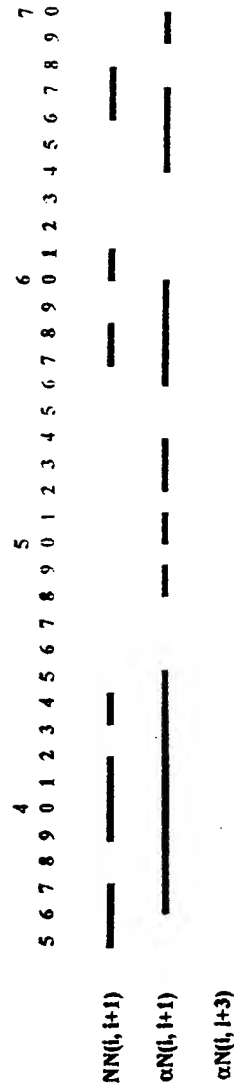
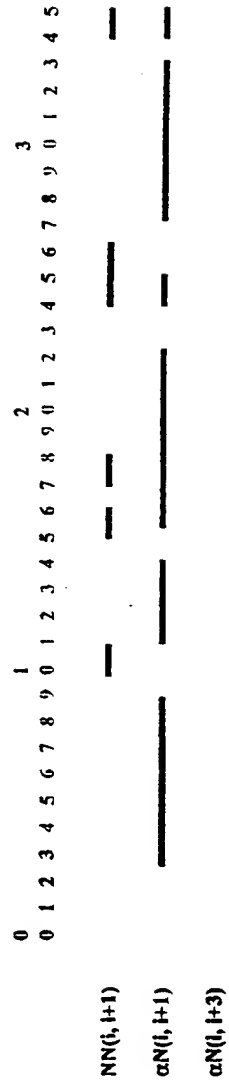


Fig 20

Cspa: Manual Assignment Analysis

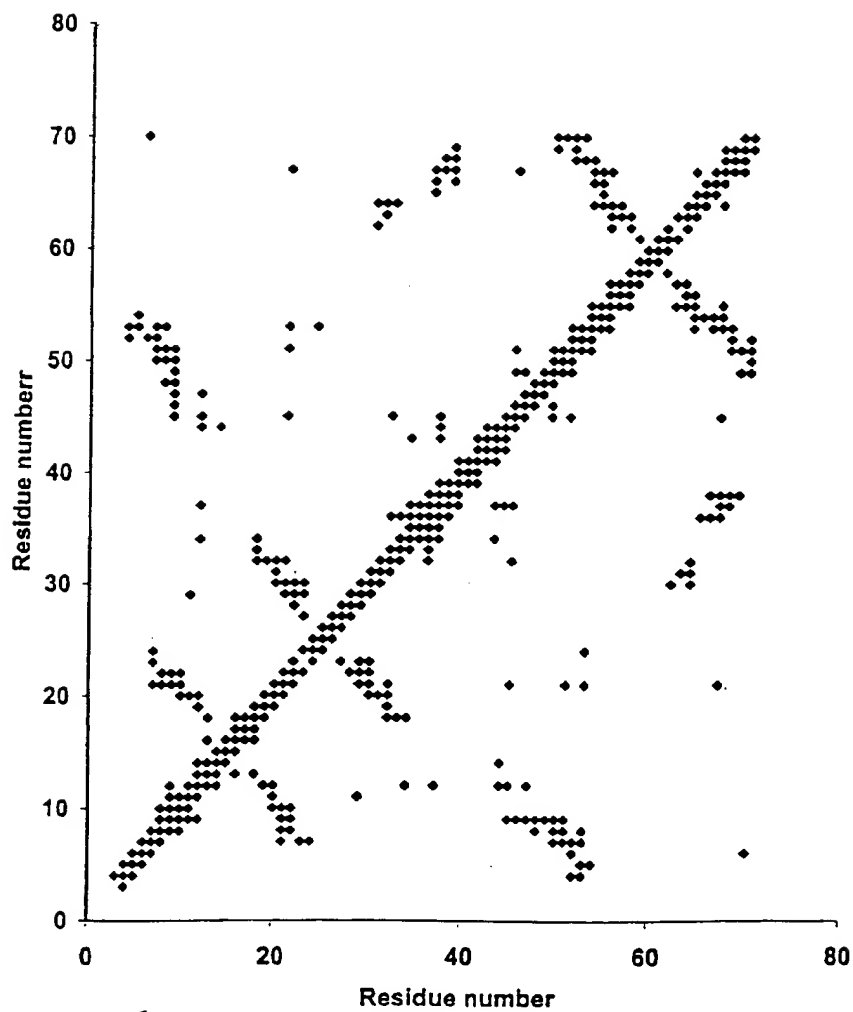
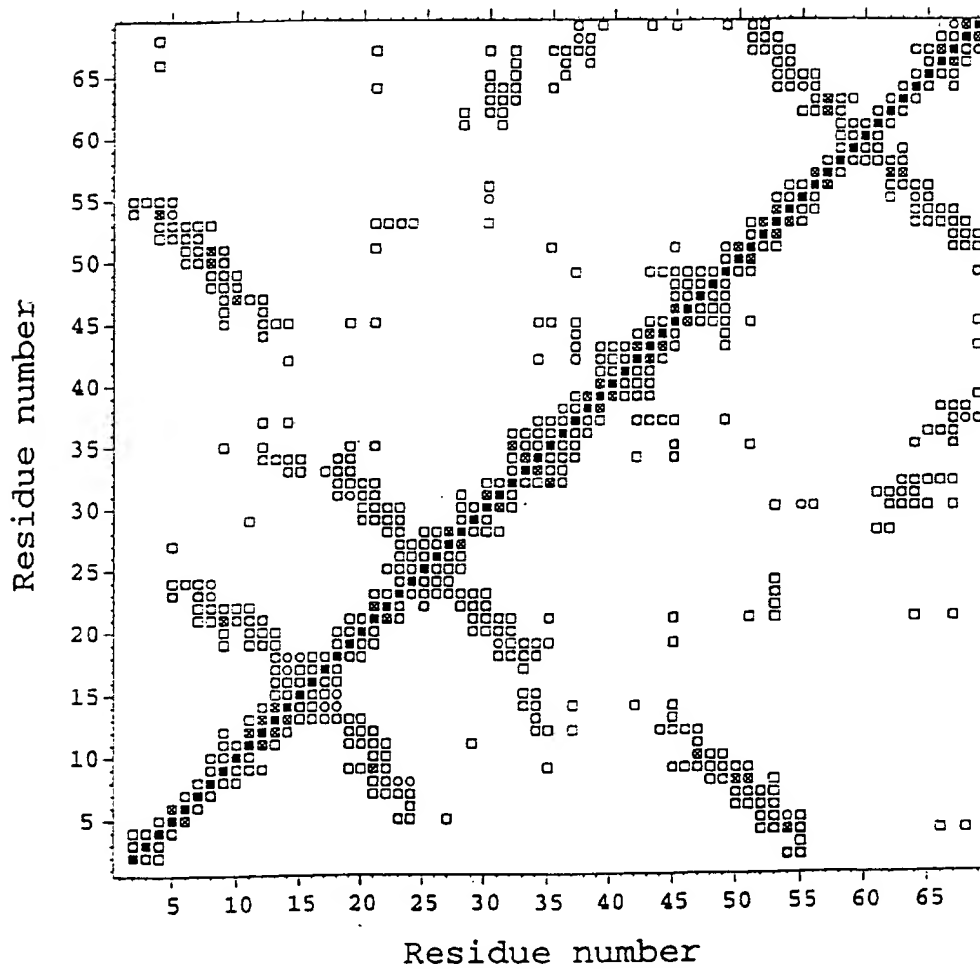


Fig 21

Cspa: Manual Structure Analysis

Fig 22

- × backbone - backbone (filled diamond if intraresidue)
- backbone - side chain (filled if intraresidue)
- side chain - side chain (filled if intraresidue)



Cspa: Manual Analysis — Connectivity Map

	1										2										3											
0	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5						
NN(l, l+1)																																
$\alpha N(l, l+1)$																																
$\alpha N(l, l+3)$																																

	4										5										6										7									
	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0	1	2	3	4	5	6	7	8	9	0				
NN(l, l+1)	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---				
$\alpha N(l, l+1)$	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---				
$\alpha N(l, l+3)$	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---	---				

Fig 23

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/16417

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C12Q 1/68; C12P 19/34; C07K 13/00; G06F 17/00

US CL : 435/6, 91.2; 530/350; 702/19, 20

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/6, 91.2; 530/350; 702/19, 20

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	WALLACE et al. Derivation of 3D coordinate templates for searching structural databases: Application to Ser-His-Asp catalytic triads in the serine proteinases and lipases. Protein Science. 1996, Vol. 5, pages 1001-1013, see entire document.	1-13
Y	FRIEDRICHS et al. An automated procedure for the assignment of protein ¹ HN, ¹⁵ N, ¹³ C ^α , ¹ H ^α , ¹³ C ^β and ¹ H ^β resonances. J. Biomol. NMR. 1994, Vol. 4, pages 703-726, see entire document.	1-13
Y	FARBER et al. Determination of eukaryotic protein coding regions using neural networks and information theory. J. Mol. Biol. 1992, Vol. 226, pages 471-479, see entire document.	1-13



Further documents are listed in the continuation of Box C.



See patent family annex.

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E earlier document published on or after the international filing date

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O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the international filing date but later than the priority date claimed

T

later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X

document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y

document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

A

document member of the same patent family

Date of the actual completion of the international search

15 NOVEMBER 1999

Date of mailing of the international search report

09 DEC 1999

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Authorized officer

JEFFREY FREDMAN

Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/16417

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	BAGBY et al. The button test: A small scale method of using microdialysis cells for assessing protein stability at concentrations suitable for NMR. J. Biomol. NMR. 1997, Vol. 10, pages 279-282, see entire document.	1-13
Y	ORENGO et al. CATH - a hierarchic classification of protein domain structures. Structure. 1997, Vol. 5, No. 8, pages 1093-1108, see entire document.	1-13

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/16417

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

WEST 1.2, CAPLUS, MEDLINE, BIOSIS, CANCERLIT, EMBASE

search terms: nuclear, magnetic, resonance, computer, algorithm, three, dimensional, protein, polypeptide, structure, function, tertiary, database, sequence